



Convention on Biological Diversity

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AD HOC TECHNICAL EXPERT GROUP ON
RISK ASSESSMENT

Online, 30 March - 3 April 2020

Item 3 of the provisional agenda*

LIST OF BIBLIOGRAPHIC REFERENCES ON ENGINEERED GENE DRIVES AND LIVING MODIFIED FISH

Note by the Executive Secretary

I. INTRODUCTION

1. In [decision CP-9/13](#), the Conference of the Parties serving as the meeting of the Parties to the Cartagena Protocol on Biosafety (COP-MOP) decided to convene the Ad Hoc Technical Expert Group (AHTEG) on risk assessment and risk management to work in accordance with the terms of reference annexed to that decision. In the same decision, the Conference of the Parties serving as the meeting of the Parties to the Cartagena Protocol requested the Executive Secretary to commission two studies informing the application of annex I to (a) living modified organisms containing engineered gene drives and (b) living modified fish and present them to the online forum and the AHTEG. The Conference of the Parties serving as the meeting of the Parties to the Cartagena Protocol also requested the Executive Secretary to collect relevant information to facilitate the work of the AHTEG.

2. Based on the above, the Secretariat has compiled a list of references from various sources with the aim to support the deliberations of the AHTEG by providing background information that may be relevant for discussion on the various agenda items.

3. The present compilation of references contains references submitted through the submissions of information on risk assessment and risk management (section II), references shared during the Open-Ended Online Forum on Risk Assessment and Risk Management (section III), as well as other additional references on the topics of engineered gene drives and living modified fish (section IV).

4. The references include peer-reviewed articles, opinion documents and perspectives.

II. LIST OF REFERENCES PROVIDED IN SUBMISSIONS OF INFORMATION

Engineered gene drives

Akbari, O. S., Bellen, H. J., Bier, E., Bullock, S. L., Burt, A., Church, G. M., Cook, K. R., Duchek, P., Edwards, O. R., Esvelt, K. M., Gantz, V. M., Golic, K. G., Gratz, S. J., Harrison, M. M., Hayes, K. R., James, A. A., Kaufman, T. C., Knoblich, J., Malik, H. S., ... Wildonger, J. (2015). Safeguarding gene drive experiments in the laboratory. *Science*, 349(6251), 927–929. <https://doi.org/10.1126/science.aac7932>

Beech, C., Vasan, S. S., Quinlan, M., Capurro, M., Alphey, L., Bayard, V., Bouaré, M., Corena-McLeod, M., Kittayapong, P., Lavery, J. V., Hanlim, L., Marrelli, M., Nagaraju, J., Ombongi, K., Othman, D. R., Pillai, V., Ramsey, J., Reuben, R., Rose, R. I., & Mumford, J. D. (2009). Deployment of innovative genetic vector control strategies: Progress on regulatory and biosafety aspects, capacity building and development of best-practice guidance. In *Asia-Pacific Journal of Molecular Biology and Biotechnology* (Vol. 17).

Benedict, M., D'Abbs, P., Dobson, S., Gottlieb, M., Harrington, L., Higgs, S., James, A., James, S., Knols, B., Lavery, J., O'Neill, S., Scott, T., Takken, W., & Toure, Y. (2008). Guidance for Contained Field Trials of Vector Mosquitoes Engineered to Contain a Gene Drive System: Recommendations of a Scientific Working Group. *Vector-Borne and Zoonotic Diseases*, 8(2), 127–166.

<https://doi.org/10.1089/vbz.2007.0273>

Benedict, M. Q., Burt, A., Capurro, M. L., De Barro, P., Handler, A. M., Hayes, K. R., Marshall, J. M., Tabachnick, W. J., & Adelman, Z. N. (2018). Recommendations for Laboratory Containment and Management of Gene Drive Systems in Arthropods. *Vector-Borne and Zoonotic Diseases*, 18(1), 2–13.

<https://doi.org/10.1089/vbz.2017.2121>

Brossard, D., Belluck, P., Gould, F., & Wirz, C. D. (2019). Promises and perils of gene drives: Navigating the communication of complex, post-normal science. *Proceedings of the National Academy of Sciences*, 116(16), 7692–7697. <https://doi.org/10.1073/pnas.1805874115>

Burt, A., & Crisanti, A. (2018). Gene Drive: Evolved and Synthetic. *ACS Chemical Biology*, 13(2), 343–346. <https://doi.org/10.1021/acschembio.7b01031>

Burt, A., & Trivers, R. (2008). *Genes in Conflict* (First). Belknap Press of Harvard University Press.

Conner, A. J., & Jacobs, J. M. E. (2019). A natural, conditional gene drive in plants. *BioRxiv*, pre-print. <https://doi.org/https://doi.org/10.1101/519884>

Cummings, C. L., & Kuzma, J. (2017). Societal Risk Evaluation Scheme (SRES): Scenario-Based Multi-Criteria Evaluation of Synthetic Biology Applications. *PLOS ONE*, 12(1), e0168564. <https://doi.org/10.1371/journal.pone.0168564>

de Jong, T. J. (2017). Gene drives do not always increase in frequency: from genetic models to risk assessment. *Journal of Consumer Protection and Food Safety*, 12(4), 299–307. <https://doi.org/10.1007/s00003-017-1131-z>

Eckhoff, P. A., Wenger, E. A., Godfray, H. C. J., & Burt, A. (2017). Impact of mosquito gene drive on malaria elimination in a computational model with explicit spatial and temporal dynamics. *Proceedings of the National Academy of Sciences*, 114(2), E255–E264. <https://doi.org/10.1073/pnas.1611064114>

Emerson, C., James, S., Littler, K., & Randazzo, F. (Fil). (2017). Principles for gene drive research. *Science*, 358(6367), 1135–1136. <https://doi.org/10.1126/science.aap9026>

Esvelt, K. M., Smidler, A. L., Catteruccia, F., & Church, G. M. (2014). Concerning RNA-guided gene drives for the alteration of wild populations. *eLife*, 3. <https://doi.org/10.7554/eLife.03401>

Gantz, V. M., Jasinskiene, N., Tatarenkova, O., Fazekas, A., Macias, V. M., Bier, E., & James, A. A. (2015). Highly efficient Cas9-mediated gene drive for population modification of the malaria vector mosquito *Anopheles stephensi*. *Proceedings of the National Academy of Sciences*, 112(49), E6736–E6743. <https://doi.org/10.1073/pnas.1521077112>

Hammond, A., Galizi, R., Kyrou, K., Simoni, A., Siniscalchi, C., Katsanos, D., Gribble, M., Baker, D., Marois, E., Russell, S., Burt, A., Windbichler, N., Crisanti, A., & Nolan, T. (2015). A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*. *Nature Biotechnology*, 34, 78. <https://doi.org/10.1038/nbt.3439>

Hayes, K. R., Hosack, G. R., Dana, G. V., Foster, S. D., Ford, J. H., Thresher, R., Ickowicz, A., Peel, D., Tizard, M., De Barro, P., Strive, T., & Dambacher, J. M. (2018). Identifying and detecting potentially adverse ecological outcomes associated with the release of gene-drive modified organisms. *Journal of Responsible Innovation*, 5(sup1), S139–S158. <https://doi.org/10.1080/23299460.2017.1415585>

Hoffmann, A. A., Montgomery, B. L., Popovici, J., Iturbe-Ormaetxe, I., Johnson, P. H., Muzzi, F., Greenfield, M., Durkan, M., Leong, Y. S., Dong, Y., Cook, H., Axford, J., Callahan, A. G., Kenny, N., Omodei, C., McGraw, E. A., Ryan, P. A., Ritchie, S. A., Turelli, M., & O'Neill, S. L. (2011). Successful establishment of Wolbachia in Aedes populations to suppress dengue transmission. *Nature*, 476(7361), 454–457. <https://doi.org/10.1038/nature10356>

James, S., Collins, F. H., Welsonhoff, P. A., Emerson, C., Godfray, H. C. J., Gottlieb, M., Greenwood, B., Lindsay, S. W., Mbogo, C. M., Okumu, F. O., Quemada, H., Savadogo, M., Singh, J. A., Tountas, K. H., & Touré, Y. T. (2018). Pathway to Deployment of Gene Drive Mosquitoes as a Potential Biocontrol Tool for Elimination of Malaria in Sub-Saharan Africa: Recommendations of a Scientific Working Group †. *The American Journal of Tropical Medicine and Hygiene*, 98(6_Suppl), 1–49. <https://doi.org/10.4269/ajtmh.18-0083>

KaramiNejadRanjbar, M., Eckermann, K. N., Ahmed, H. M. M., Sánchez C., H. M., Dippel, S., Marshall, J. M., & Wimmer, E. A. (2018). Consequences of resistance evolution in a Cas9-based sex conversion-suppression gene drive for insect pest management. *Proceedings of the National Academy of Sciences*, 115(24), 6189–6194. <https://doi.org/10.1073/pnas.1713825115>

Lindholm, A. K., Dyer, K. A., Firman, R. C., Fishman, L., Forstmeier, W., Holman, L., Johannesson, H., Knief, U., Kokko, H., Larracuente, A. M., Manser, A., Montchamp-Moreau, C., Petrosyan, V. G., Pomiankowski, A., Presgraves, D. C., Safronova, L. D., Sutter, A., Unckless, R. L., Verspoor, R. L., ... Price, T. A. R. (2016). The Ecology and Evolutionary Dynamics of Meiotic Drive. *Trends in Ecology & Evolution*, 31(4), 315–326. <https://doi.org/10.1016/j.tree.2016.02.001>

Lunshof, J. E., & Birnbaum, A. (2017). Adaptive Risk Management of Gene Drive Experiments. *Applied Biosafety*, 22(3), 97–103. <https://doi.org/10.1177/1535676017721488>

Meghani, Z., & Kuzma, J. (2018). Regulating animals with gene drive systems: lessons from the regulatory assessment of a genetically engineered mosquito. *Journal of Responsible Innovation*, 5(sup1), S203–S222. <https://doi.org/10.1080/23299460.2017.1407912>

Noble, C., Adlam, B., Church, G. M., Esvelt, K. M., & Nowak, M. A. (2018). Current CRISPR gene drive systems are likely to be highly invasive in wild populations. *eLife*, 7. <https://doi.org/10.7554/eLife.33423>

Oye, K. A., Esvelt, K., Appleton, E., Catteruccia, F., Church, G., Kuiken, T., Lightfoot, S. B.-Y., McNamara, J., Smidler, A., & Collins, J. P. (2014). Regulating gene drives. *Science*, 345(6197), 626–628. <https://doi.org/10.1126/science.1254287>

Oye, K. (2014, August). Proceed with Caution. *MIT Technology Review*. <https://www.technologyreview.com/s/530156/proceed-with-caution/>

Roberts, A., Andrade, P. P. de, Okumu, F., Quemada, H., Savadogo, M., Singh, J. A., & James, S. (2017). Results from the Workshop “Problem Formulation for the Use of Gene Drive in Mosquitoes.” *The American Journal of Tropical Medicine and Hygiene*, 96(3), 530–533. <https://doi.org/10.4269/ajtmh.16-0726>

Roberts, A., Andrade, P. P. de, Okumu, F., Quemada, H., Savadogo, M., Singh, J. A., & James, S. (2017). Results from the Workshop “Problem Formulation for the Use of Gene Drive in Mosquitoes.” *The American Journal of Tropical Medicine and Hygiene*, 96(3), 530–533. <https://doi.org/10.4269/ajtmh.16-0726>

Rudenko, L., Palmer, M. J., & Oye, K. (2018). Considerations for the governance of gene drive organisms. *Pathogens and Global Health*, 112(4), 162–181. <https://doi.org/10.1080/20477724.2018.1478776>

Sandler, L., Hiraizumi, Y., & Sandler, I. (1959). Meiotic Drive in Natural Populations of *Drosophila melanogaster*. I. the Cytogenetic Basis of Segregation-Distortion. *Genetics*, 44(2), 233–250. <https://www.genetics.org/content/44/2/233.long>

Simon, S., Otto, M., & Engelhard, M. (2018). Synthetic gene drive: between continuity and novelty. *EMBO Reports*, 19(5), e45760. <https://doi.org/10.15252/embr.201845760>

van der Vlugt, C. J. B., Brown, D. D., Lehmann, K., Leunda, A., & Willemarck, N. (2018). A Framework for the Risk Assessment and Management of Gene Drive Technology in Contained Use. *Applied Biosafety*, 23(1), 25–31. <https://doi.org/10.1177/1535676018755117>

Werren, J. H., Nur, U., & Wu, C.-I. (1988). Selfish genetic elements. *Trends in Ecology & Evolution*, 3(11), 297–302. [https://doi.org/10.1016/0169-5347\(88\)90105-X](https://doi.org/10.1016/0169-5347(88)90105-X)

Living modified fish

Abrahams, M. V., & Sutterlin, A. (1999). The foraging and antipredator behaviour of growth-enhanced transgenic Atlantic salmon. *Animal Behaviour*, 58(5), 933–942. <https://doi.org/10.1006/anbe.1999.1229>

Cnaani, A., McLean, E., & Hallerman, E. M. (2013). Effects of growth hormone transgene expression and triploidy on acute stress indicators in Atlantic salmon (*Salmo salar* L.). *Aquaculture*, 412–413, 107–116. <https://doi.org/10.1016/j.aquaculture.2013.06.029>

Cook, J. T., McNiven, M. A., & Sutterlin, A. M. (2000). Metabolic rate of pre-smolt growth-enhanced transgenic Atlantic salmon (*Salmo salar*). *Aquaculture*, 188(1–2), 33–45. [https://doi.org/10.1016/S0044-8486\(00\)00332-X](https://doi.org/10.1016/S0044-8486(00)00332-X)

Cook, J. ., McNiven, M. ., Richardson, G. ., & Sutterlin, A. . (2000). Growth rate, body composition and feed digestibility/conversion of growth-enhanced transgenic Atlantic salmon (*Salmo salar*). *Aquaculture*, 188(1–2), 15–32. [https://doi.org/10.1016/S0044-8486\(00\)00331-8](https://doi.org/10.1016/S0044-8486(00)00331-8)

Cook, J. ., Sutterlin, A. ., & McNiven, M. . (2000). Effect of food deprivation on oxygen consumption and body composition of growth-enhanced transgenic Atlantic salmon (*Salmo salar*). *Aquaculture*, 188(1–2), 47–63. [https://doi.org/10.1016/S0044-8486\(00\)00333-1](https://doi.org/10.1016/S0044-8486(00)00333-1)

Cotter, D., O’Donovan, V., O’Maoiléidigh, N., Rogan, G., Roche, N., & Wilkins, N. P. (2000). An evaluation of the use of triploid Atlantic salmon (*Salmo salar* L.) in minimising the impact of escaped farmed salmon on wild populations. *Aquaculture*, 186(1–2), 61–75. [https://doi.org/10.1016/S0044-8486\(99\)00367-1](https://doi.org/10.1016/S0044-8486(99)00367-1)

Crossin, G. T., & Devlin, R. H. (2017). Predation, metabolic priming and early life-history rearing environment affect the swimming capabilities of growth hormone transgenic rainbow trout. *Biology Letters*, 13(8), 20170279. <https://doi.org/10.1098/rsbl.2017.0279>

Crossin, G. T., Sundström, L. F., Vandersteen, W. E., & Devlin, R. H. (2015). Early Life-History Consequences of Growth-Hormone Transgenesis in Rainbow Trout Reared in Stream Ecosystem Mesocosms. *PLOS ONE*, 10(3), e0120173. <https://doi.org/10.1371/journal.pone.0120173>

Deitch, E. J. (2006). Cardiorespiratory modifications, and limitations, in post-smolt growth hormone transgenic Atlantic salmon *Salmo salar*. *Journal of Experimental Biology*, 209(7), 1310–1325. <https://doi.org/10.1242/jeb.02105>

Devlin, R. H., D'Andrade, M., Uh, M., & Biagi, C. A. (2004). Population effects of growth hormone transgenic coho salmon depend on food availability and genotype by environment interactions. *Proceedings of the National Academy of Sciences*, 101(25), 9303–9308. <https://doi.org/10.1073/pnas.0400023101>

Devlin, R. H., Sundström, L. F., & Muir, W. M. (2006). Interface of biotechnology and ecology for environmental risk assessments of transgenic fish. *Trends in Biotechnology*, 24(2), 89–97. <https://doi.org/10.1016/j.tibtech.2005.12.008>

Devlin, R. H., Yesaki, T. Y., Donaldson, E. M., Du, S. J., & Hew, C.-L. (1995). Production of germline transgenic Pacific salmonids with dramatically increased growth performance. *Canadian Journal of Fisheries and Aquatic Sciences*, 52(7), 1376–1384. <https://doi.org/10.1139/f95-133>

Devlin, R. H., Yesaki, T. Y., Donaldson, E. M., & Hew, C.-L. (1995). Transmission and phenotypic effects of an antifreeze/GH gene construct in coho salmon (*Oncorhynchus kisutch*). *Aquaculture*, 137(1–4), 161–169. [https://doi.org/10.1016/0044-8486\(95\)01090-4](https://doi.org/10.1016/0044-8486(95)01090-4)

Devlin, R. H., Biagi, C. A., & Yesaki, T. Y. (2004). Growth, viability and genetic characteristics of GH transgenic coho salmon strains. *Aquaculture*, 236(1–4), 607–632. <https://doi.org/10.1016/j.aquaculture.2004.02.026>

Down, N. E., Donaldson, E. M., Dye, H. M., Boone, T. C., Langley, K. E., & Souza, L. M. (1989). A Potent Analog of Recombinant Bovine Somatotropin Accelerates Growth in Juvenile Coho Salmon (*Oncorhynchus kisutch*). *Canadian Journal of Fisheries and Aquatic Sciences*, 46(2), 178–183. <https://doi.org/10.1139/f89-024>

Du, S. J., Gong, Z., Fletcher, G. L., Shears, M. A., King, M. J., Idler, D. R., & Hew, C. L. (1992). Growth Enhancement in Transgenic Atlantic Salmon by the Use of an “All Fish” Chimeric Growth Hormone Gene Construct. *Nature Biotechnology*, 10(2), 176–181. <https://doi.org/10.1038/nbt0292-176>

Elliott, J. M. (1991). Tolerance and resistance to thermal stress in juvenile Atlantic salmon, *Salmo salar*. *Freshwater Biology*, 25(1), 61–70. <https://doi.org/10.1111/j.1365-2427.1991.tb00473.x>

Frankham, R. (1995). Conservation Genetics. *Annual Review of Genetics*, 29(1), 305–327. <https://doi.org/10.1146/annurev.ge.29.120195.001513>

Fredrik Sundstrom, L., Lohmus, M., Devlin, R. H., Johnsson, J. I., Biagi, C. A., & Bohlin, T. (2004). Feeding on Profitable and Unprofitable Prey: Comparing Behaviour of Growth-Enhanced Transgenic and

Normal Coho Salmon (*Oncorhynchus kisutch*). *Ethology*, 110(5), 381–396. <https://doi.org/10.1111/j.1439-0310.2004.00985.x>

Gjøen, H., & Bentsen, H. B. (1997). Past, present, and future of genetic improvement in salmon aquaculture. *ICES Journal of Marine Science*, 54(6), 1009–1014. [https://doi.org/10.1016/S1054-3139\(97\)80005-7](https://doi.org/10.1016/S1054-3139(97)80005-7)

Gray, A. K., Evans, M. A., & Thorgaard, G. H. (1993). Viability and development of diploid and triploid salmonid hybrids. *Aquaculture*, 112(2–3), 125–142. [https://doi.org/10.1016/0044-8486\(93\)90439-6](https://doi.org/10.1016/0044-8486(93)90439-6)

Hallerman, E. M., McLean, E., & Fleming, I. A. (2007). Effects of growth hormone transgenes on the behavior and welfare of aquacultured fishes: A review identifying research needs. *Applied Animal Behaviour Science*, 104(3–4), 265–294. <https://doi.org/10.1016/j.applanim.2006.09.008>

Jansson, H., Holmgren, I., Wedin, K., & Anderson, T. (1991). High frequency of natural hybrids between Atlantic salmon, *Salmo salar* L., and brown trout, *S. trutta* L., in a Swedish river. *Journal of Fish Biology*, 39, 343–348. <https://doi.org/10.1111/j.1095-8649.1991.tb05096.x>

Johnsson, J. I., & Bjornsson, B. T. (2001). Growth-enhanced fish can be competitive in the wild. *Functional Ecology*, 15(5), 654–659. <https://doi.org/10.1046/j.0269-8463.2001.00566.x>

KAPUSCINSKI, A. R. (2005). Estado de los conocimientos científicos sobre el nivel de bioseguridad para el medio ambiente que presentan los peces y crustáceos transgénicos. *Revue Scientifique et Technique de l'OIE*, 24(1), 309–322. <https://doi.org/10.20506/rst.24.1.1576>

Kapuscinski, A. R., & Brister, D. J. (2001). Genetic impacts of aquaculture. In K. D. Black (Ed.), *Environmental Impacts of Aquaculture* (pp. 385–415). Sheffield Academic Press.

Kapuscinski, A. R., & Hallerman, E. M. (1990). Transgenic Fish and Public Policy: Anticipating Environmental Impacts of Transgenic Fish. *Fisheries*, 15(1), 2–11. [https://doi.org/10.1577/1548-8446\(1990\)015<0002:TFAPPA>2.0.CO;2](https://doi.org/10.1577/1548-8446(1990)015<0002:TFAPPA>2.0.CO;2)

Kapuscinski, A. R., & Hallerman, E. M. (1991). Implications of Introduction of Transgenic Fish into Natural Ecosystems. *Canadian Journal of Fisheries and Aquatic Sciences*, 48(S1), 99–107. <https://doi.org/10.1139/f91-308>

Kapuscinski, A. R., Hard, J. J., Paulson, K. M., Neira, R., Ponniah, A., Kamonrat, W., Mwanja, W., Fleming, I. A., Gallardo, J., Devlin, R. H., & Trisak, J. (2007). Approaches to assessing gene flow. In A. R. Kapuscinski, S. Li, K. R. Hayes, G. V. Dana, E. M. Hallerman, & P. J. Schei (Eds.), *Environmental Risk Assessment of Genetically Modified Organisms Volume 3. Methodologies for Transgenic Fish* (pp. 1–304). CAB International.

King, T. L., Kalinowski, S. T., Schill, W. B., Spidle, A. P., & Lubinski, B. A. (2001). Population structure of Atlantic salmon (*Salmo salar* L.): a range-wide perspective from microsatellite DNA variation. *Molecular Ecology*, 10(4), 807–821. <https://doi.org/10.1046/j.1365-294X.2001.01231.x>

Leggatt, R. A., Biagi, C. A., Sakhrani, D., Dominelli, R., Eliason, E. J., Farrell, A. P., & Devlin, R. H. (2017). Fitness component assessments of wild-type and growth hormone transgenic coho salmon reared in seawater mesocosms. *Aquaculture*, 473, 31–42. <https://doi.org/10.1016/j.aquaculture.2017.01.022>

Leggatt, R. A., Sundström, L. F., Woodward, K., & Devlin, R. H. (2017). Growth-Enhanced Transgenic Coho Salmon (*Oncorhynchus kisutch*) Strains Have Varied Success in Simulated Streams: Implications for Risk Assessment. *PLOS ONE*, 12(1), e0169991. <https://doi.org/10.1371/journal.pone.0169991>

Löhmus, M., Sundström, L. F., Björklund, M., & Devlin, R. H. (2010). Genotype-Temperature Interaction in the Regulation of Development, Growth, and Morphometrics in Wild-Type, and Growth-Hormone Transgenic Coho Salmon. *PLoS ONE*, 5(4), e9980. <https://doi.org/10.1371/journal.pone.0009980>

Mair, G. C., Nam, Y. K., & Solar, I. . (2007). Risk Management: reducing risk through confinement of transgenic fish. In A. R. Kapuscinski, S. Li, K. R. Hayes, G. V. Dana, E. M. Hallerman, & P. J. Schei (Eds.), *Environmental Risk Assessment of Genetically Modified Organisms Volume 3. Methodologies for Transgenic Fish*. CAB International.

Martinez-Porcha, M., & Martinez-Cordova, L. R. (2012). World Aquaculture: Environmental Impacts and Troubleshooting Alternatives. *The Scientific World Journal*, 2012, 1–9.
<https://doi.org/10.1100/2012/389623>

McCormick, S. D., Saunders, R. L., Henderson, E. B., & Harmon, P. R. (1987). Photoperiod Control of Parr-Smolt Transformation in Atlantic Salmon (*Salmo salar*): Changes in Salinity Tolerance, Gill Na⁺,K⁺-ATPase Activity, and Plasma Thyroid Hormones. *Canadian Journal of Fisheries and Aquatic Sciences*, 44(8), 1462–1468. <https://doi.org/10.1139/f87-175>

McGinnity, P. (1997). Genetic impact of escaped farmed Atlantic salmon (*Salmo salar* L.) on native populations: use of DNA profiling to assess freshwater performance of wild, farmed, and hybrid progeny in a natural river environment. *ICES Journal of Marine Science*, 54(6), 998–1008.
[https://doi.org/10.1016/S1054-3139\(97\)80004-5](https://doi.org/10.1016/S1054-3139(97)80004-5)

Moreau, D. T. R., Fleming, I. A., Fletcher, G. L., & Brown, J. A. (2011). Growth hormone transgenesis does not influence territorial dominance or growth and survival of first-feeding Atlantic salmon *Salmo salar* in food-limited stream microcosms. *Journal of Fish Biology*, 78(3), 726–740.
<https://doi.org/10.1111/j.1095-8649.2010.02888.x>

Moreau, D. T. R., Conway, C., & Fleming, I. A. (2011). Reproductive performance of alternative male phenotypes of growth hormone transgenic Atlantic salmon (*Salmo salar*). *Evolutionary Applications*, 4(6), 736–748. <https://doi.org/10.1111/j.1752-4571.2011.00196.x>

Moreau, D. T. R., & Fleming, I. A. (2012). Enhanced growth reduces precocial male maturation in Atlantic salmon. *Functional Ecology*, 26(2), 399–405. <https://doi.org/10.1111/j.1365-2435.2011.01941.x>

Moreau, D. T. R., Gamperl, A. K., Fletcher, G. L., & Fleming, I. A. (2014). Delayed Phenotypic Expression of Growth Hormone Transgenesis during Early Ontogeny in Atlantic Salmon (*Salmo salar*)? *PLoS ONE*, 9(4), e95853. <https://doi.org/10.1371/journal.pone.0095853>

Moreau, D. T. R. (2011). *Potential for ecological effects and gene flow resulting from growth hormone transgenic Atlantic salmon (*Salmo salar*) interactions with wild specific*. Memorial University of Newfoundland.

Moreau, D. T. R. (2014). Ecological Risk Analysis and Genetically Modified Salmon: Management in the Face of Uncertainty. *Annual Review of Animal Biosciences*, 2(1), 515–533.
<https://doi.org/10.1146/annurev-animal-022513-114231>

Mori, T., & Devlin, R. H. (1999). Transgene and host growth hormone gene expression in pituitary and nonpituitary tissues of normal and growth hormone transgenic salmon. *Molecular and Cellular Endocrinology*, 149(1–2), 129–139. [https://doi.org/10.1016/S0303-7207\(98\)00248-2](https://doi.org/10.1016/S0303-7207(98)00248-2)

Muir, W. M. (2004). The threats and benefits of GM fish. *EMBO Reports*, 5(7), 654–659. <https://doi.org/10.1038/sj.embo.7400197>

Oke, K. B., Westley, P. A. H., Moreau, D. T. R., & Fleming, I. A. (2013). Hybridization between genetically modified Atlantic salmon and wild brown trout reveals novel ecological interactions. *Proceedings of the Royal Society B: Biological Sciences*, 280(1763), 20131047–20131047. <https://doi.org/10.1098/rspb.2013.1047>

Polymeropoulos, E. T., Plouffe, D., LeBlanc, S., Elliott, N. G., Currie, S., & Frappell, P. B. (2014). Growth hormone transgenesis and polyploidy increase metabolic rate, alter the cardiorespiratory response and influence HSP expression in response to acute hypoxia in Atlantic salmon (*Salmo salar*) yolk-sac alevins. *Journal of Experimental Biology*, 217(13), 2268–2276. <https://doi.org/10.1242/jeb.098913>

Raven, P. A., Devlin, R. H., & Higgs, D. A. (2006). Influence of dietary digestible energy content on growth, protein and energy utilization and body composition of growth hormone transgenic and non-transgenic coho salmon (*Oncorhynchus kisutch*). *Aquaculture*, 254(1–4), 730–747. <https://doi.org/10.1016/j.aquaculture.2005.11.009>

Saunders, R. L., Fletcher, G. L., & Hew, C. L. (1998). Smolt development in growth hormone transgenic Atlantic salmon. *Aquaculture*, 168(1–4), 177–193. [https://doi.org/10.1016/S0044-8486\(98\)00348-2](https://doi.org/10.1016/S0044-8486(98)00348-2)

Snow, A. A., Andow, D. A., Gepts, P., Hallerman, E. M., Power, A., Tiedje, J. M., & Wolfenbarger, L. L. (2005). Genetically engineered organisms and the environment: Current Status and recommendations 1. *Ecological Applications*, 15(2), 377–404. <https://doi.org/10.1890/04-0539>

Ståhl, G. (1987). Genetic population structure of Atlantic salmon. In N. Ryman & F. Utter (Eds.), *Population Genetics and Fishery Management* (pp. 121–140). University of Washington Press.

Stead, S. M., & Laird, L. (2002). *The Handbook of Salmon Farming* (E. D. Stevens & A. Stevens (Eds.); 978th-1st–8523rd ed.). Springer-Verlag London, Praxis Publishing.

Stevens, E. D., & Sutterlin, A. (1999). Gill Morphometry in Growth Hormone Transgenic Atlantic Salmon. *Environmental Biology of Fishes*, 54(4), 405–411. <https://doi.org/10.1023/A:1007574308129>

Stevens, E. D., Sutterlin, A., & Cook, T. (1998). Respiratory metabolism and swimming performance in growth hormone transgenic Atlantic salmon. *Canadian Journal of Fisheries and Aquatic Sciences*, 55(9), 2028–2035. <https://doi.org/10.1139/f98-078>

Sundström, L. F., Löhmus, M., & Devlin, R. H. (2016). Gene-environment interactions influence feeding and anti-predator behavior in wild and transgenic coho salmon. *Ecological Applications*, 26(1), 67–76. <https://doi.org/10.1890/15-0252>

Sundstrom, L. F., Lohmus, M., Tymchuk, W. E., & Devlin, R. H. (2007). Gene-environment interactions influence ecological consequences of transgenic animals. *Proceedings of the National Academy of Sciences*, 104(10), 3889–3894. <https://doi.org/10.1073/pnas.0608767104>

Tibbetts, S. M., Wall, C. L., Barbosa-Solomieu, V., Bryenton, M. D., Plouffe, D. A., Buchanan, J. T., & Lall, S. P. (2013). Effects of combined 'all-fish' growth hormone transgenics and triploidy on growth and nutrient utilization of Atlantic salmon (*Salmo salar* L.) fed a practical grower diet of known composition. *Aquaculture*, 406–407, 141–152. <https://doi.org/10.1016/j.aquaculture.2013.05.005>

Wong, A. C., & Van Eenennaam, A. L. (2008). Transgenic approaches for the reproductive containment of genetically engineered fish. *Aquaculture*, 275(1–4), 1–12. <https://doi.org/10.1016/j.aquaculture.2007.12.026>

Relevant reports and resources

Agricultural Biotechnology Research Advisory Committee. (1995). *Performance standards for safely conducting research with genetically modified fish and shellfish. Document No. 95-04.*

Australian Academy of Science. (2017). *Synthetic gene drives in Australia: Implications of emerging technologies.* <http://www.science.org.au/gene-drives>

Bioteknologirådet (The Norwegian Biotechnology Advisory Board). (2017). *Statement on Gene Drives.* <http://www.bioteknologiradet.no/filarkiv/2017/02/Statement-on-gene-drives.pdf>

Buchori, D., Aryati, S., Hadi, U. K., & Joseph, H. K. (2017). *Risk Assessment on the Release of Wolbachia-infected Aedes aegypti.*

<http://www.eliminatedengue.com/yogyakarta/download/view/publication/379>

Department of Health Office of the Gene Technology Regulator. (2016). *Guidance for IBCs: Regulatory requirements for contained research with GMOs containing engineered gene drives.* <http://www.ogtr.gov.au/internet/ogtr/publishing.nsf/Content/ibc-1>

EFSA. (2016). *Guidelines on Active Substances and Plant Protection Products.*

https://ec.europa.eu/food/plant/pesticides/approval_active_substances/guidance_documents_en

EFSA GMO Panel. (2013). Guidance on the environmental risk assessment of genetically modified animals. *EFSA Journal*, 11(5), 3200. <https://doi.org/10.2903/j.efsa.2013.3200>

EFSA Panel on Genetically Modified Organisms. (2013). Guidance on the environmental risk assessment of genetically modified animals. *EFSA Journal*, 11(5), 3200. <https://doi.org/10.2903/j.efsa.2013.3200>

EFSA Panels on GMO and AHAW. (2012). Guidance on the risk assessment of food and feed from genetically modified animals and on animal health and welfare aspects. *EFSA Journal*, 10(1), 2501. <https://doi.org/10.2903/j.efsa.2012.2501>

Directive 2001/18/EC of the European Parliament and Council of 12 March 2001 on the deliberate release into the environment of genetically modified organisms and repealing Council Directive 90/220/EEC, 61 (2018) (testimony of European Parliament). <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:02001L0018-20180329&qid=1550847942646&from=EN>

European Union. (2009). *Directive 2009/41/EC of the European Parliament and the Council of 6 May 2009 on the contained use of genetically modified micro-organisms.*

Hindar, K. (1993). *Genetically engineered fish and their possible environmental impact.*

International Council for the Exploration of the Sea. (2010). *ICES Annual Report 2009*.

Jeger, M., Bragard, C., Caffier, D., Candresse, T., Chatzivassiliou, E., Dehnen-Schmutz, K., Grégoire, J., Jaques Miret, J. A., MacLeod, A., Navajas Navarro, M., Niere, B., Parnell, S., Potting, R., Rafoss, T., Rossi, V., Urek, G., Van Bruggen, A., Van Der Werf, W., West, J., ... Gilioli, G. (2018). Guidance on quantitative pest risk assessment. *EFSA Journal*, 16(8). <https://doi.org/10.2903/j.efsa.2018.5350>

Murphy, B., Jansen, C., Murray, J., & De Barro, P. (2010). *Risk Analysis on the Australian release of Aedes aegypti (L.) (Diptera: Culicidae) containing Wolbachia*.

National Academies of Sciences Engineering and Medicine. (2016). *Gene Drives on the Horizon*. National Academies Press. <https://doi.org/10.17226/23405>

National Research Council. (2004). *Biological Confinement of Genetically Engineered Organisms*. National Academies Press. <https://doi.org/10.17226/10880>

OECD. (2019). *Safety of novel foods and feeds and on the harmonisation of regulatory oversight in biotechnology*.

<http://www.oecd.org/chemicalsafety/biotrack/oecdandrisksafetyassessmentinmodernbiotechnology.htm>

OECD. (2017). *Consensus document on the biology of atlantic salmon (Salmo satar)*.

<https://doi.org/https://doi.org/10.1787/9789264279728-en>

Oficina Internacional del Trabajo. (2014). *Convenio Núm. 169 de la OIT sobre Pueblos Indígenas y Tribales: Declaración de las Naciones Unidas sobre los Derechos de los Pueblos Indígenas*.

https://www.ilo.org/wcmsp5/groups/public/---americas/---ro-lima/documents/publication/wcms_345065.pdf

Scientific Advice Mechanism High Level Group of Scientific Advisors. (2017). *New techniques in agricultural biotechnology*. European Commission.

<https://ec.europa.eu/research/sam/index.cfm?pg=agribiotechnology>

Scientific Committee of the Haut Conseil des Biotechnologies. (2014). *Scientific Opinion in response to the referral of 12 October 2015 concerning use of genetically modified mosquitoes for vector control*.

<http://www.hautconseildesbiotechnologies.fr/>

Scientific Committee on Health Environmental and Emerging Risks. (2018). *Statement on emerging health and environmental issues (2018)*. <https://doi.org/10.2875/840266>

Scientific Committee on Health, E. and E. R. (2018). *Emerging Issues and the Role of the SCHEER*. <https://doi.org/10.2875/33037>

Secretariat of the International Plant Protection Convention. (2009). *Training material on pest risk analysis based on IPPC standards*. <https://www.ippc.int/en/core-activities/capacity-development/training-material-pest-risk-analysis-based-ippc-standards/>

Secretariat of the International Plant Protection Convention. (2006). *International Standards for Phytosanitary Measures: 1 to 24*. <http://www.fao.org/3/a0450e/a0450e00.htm>

Teufel, J., Pätzold, F., & Potthof, C. (2002). *Specific research on transgenic fish considering especially the biology of trout and salmon.*

U.S. Department of Health and Human Services. (2019). *NIH Guidelines for Research involving recombinant or synthetic nucleic acid molecules (NIH Guidelines).* https://osp.od.nih.gov/wp-content/uploads/NIH_Guidelines.html

U.S. Fish & Wildlife Service. (2009). *Altantic salmon (Salmo salar).* <https://ecos.fws.gov/ecp0/profile/speciesProfile?spcode=E07L>

United States Environmental Protection Agency. (1992). *Framework for Ecological Risk Assessment.* <https://www.epa.gov/risk/framework-ecological-risk-assessment>

United States Environmental Protection Agency. (2019). *Risk Assessment Guidelines.* <https://www.epa.gov/risk/risk-assessment-guidelines>

United States Environmental Protection Agency, United States Food and Drug Administration, & United States Department of Agriculture. (2017). *Modernizing the Regulatory System for Biotechnology Products: Final Version of the 2017 Update to the Coordinated Framework for the Regulation of Biotechnology.* https://www.aphis.usda.gov/biotechnology/downloads/2017_coordinated_framework_update.pdf

Westra, J., van der Vlugt, C. J. B., Roesink, C., Hogervorst, P. A. M., & Glandorf, D. C. M. (2016). *Genes Drives: Policy Report.* <https://rivm.openrepository.com/handle/10029/596002>

World Health Organization. (2015). *World Malaria Report 2015.* <https://www.who.int/malaria/publications/world-malaria-report-2015/report/en/>

World Health Organization/Special Programme for Research and Training in Tropical Diseases, & Foundation for the National Institutes of Health. (2014). *Guidance framework for testing of genetically modified mosquitoes.* <https://www.who.int/tdr/publications/year/2014/guide-fmrk-gm-mosquit/en/>

World Organisation for Animal Health. (2019). *Invasive alien animal species.* <http://www.oie.int/en/our-scientific-expertise/specific-information-and-recommendations/invasive-alien-animal-species/>

Zentrale Kommission für die Biologische Sicherheit. (2016). *Position statement of the ZKBS on the classification of genetic engineering operations for the production and use of higher organisms using recombinant gene drive systems.* 1–4.

Zentrale Kommission für die Biologische Sicherheit. (2011). *Preliminary assessment of the ZKBS of the available documents regarding an application by the company Aqua Bounty Technologies for approval of genetically modified salmon (brand name AquAdvantage) in the U.S. in respect of potential environmental risks.*

III. LIST OF REFERENCE CITED DURING THE OPEN-ENDED ONLINE FORUM ON RISK ASSESSMENT AND RISK MANAGEMENT

Topic 1: Engineered gene drives

Publications, reports and communications

Adelman, Z. N., Pledger, D., & Myles, K. M. (2017). Developing standard operating procedures for gene drive research in disease vector mosquitoes. *Pathogens and Global Health*, 111(8), 436–447. <https://doi.org/10.1080/20477724.2018.1424514>

African Centre for Biodiversity. (2018). Critique of African Union and NEPAD's positions on gene drive mosquitoes for Malaria elimination (p. 18).

https://www.acbio.org.za/sites/default/files/documents/Critique_of_African_Union_and_NEPADs_positions_on_gene_drive_mosquitoes_for_Malaria_elimination.pdf

African Union, & NEPAD. (2018). Gene drives for malaria control and elimination in Africa.

www.nepad.org

Ågren, J. A., & Clark, A. G. (2018). Selfish genetic elements. PLOS Genetics, 14(11), e1007700.

<https://doi.org/10.1371/journal.pgen.1007700>

Akbari, O. S., Bellen, H. J., Bier, E., Bullock, S. L., Burt, A., Church, G. M., Cook, K. R., Duchek, P., Edwards, O. R., Esveld, K. M., Gantz, V. M., Golic, K. G., Gratz, S. J., Harrison, M. M., Hayes, K. R., James, A. A., Kaufman, T. C., Knoblich, J., Malik, H. S., ... Wildonger, J. (2015). Safeguarding gene drive experiments in the laboratory. Science, 349(6251), 927–929. <https://doi.org/10.1126/science.aac7932>

Akbari, O. S., Antoshechkin, I., Amrhein, H., Williams, B., Diloreto, R., Sandler, J., & Hay, B. A. (2013). The Developmental Transcriptome of the Mosquito *Aedes aegypti*, an Invasive Species and Major Arbovirus Vector. G3: Genes|Genomes|Genetics, 3(9), 1493–1509. <https://doi.org/10.1534/g3.113.006742>

Akbari, O. S., Matzen, K. D., Marshall, J. M., Huang, H., Ward, C. M., & Hay, B. A. (2013). A Synthetic Gene Drive System for Local, Reversible Modification and Suppression of Insect Populations. Current Biology, 23(8), 671–677. <https://doi.org/10.1016/j.cub.2013.02.059>

Ali, M. W., Zheng, W., Sohail, S., Li, Q., Zheng, W., & Zhang, H. (2017). A genetically enhanced sterile insect technique against the fruit fly, *Bactrocera dorsalis* (Hendel) by feeding adult double-stranded RNAs. Scientific Reports, 7(1), 4063. <https://doi.org/10.1038/s41598-017-04431-z>

Alphey, L. (2014). Genetic Control of Mosquitoes. Annual Review of Entomology, 59(1), 205–224. <https://doi.org/10.1146/annurev-ento-011613-162002>

Anderson, M. E., Mavica, J., Shackleford, L., Flis, I., Fochler, S., Basu, S., & Alphey, L. (2019). CRISPR/Cas9 gene editing in the West Nile Virus vector, *Culex quinquefasciatus* Say. PLOS ONE, 14(11), e0224857. <https://doi.org/10.1371/journal.pone.0224857>

Australian Government Department of Health. (2019). Guidance for IBCs: Regulatory requirements for contained research with GMOs containing engineered gene drives (p. 2).

[http://www.ogtr.gov.au/internet/ogtr/publishing.nsf/Content/53139D205A98A3B3CA257D4F00811F97\\$/File/Guidance_on_gene_drives.pdf](http://www.ogtr.gov.au/internet/ogtr/publishing.nsf/Content/53139D205A98A3B3CA257D4F00811F97$/File/Guidance_on_gene_drives.pdf)

Bae, S., Park, J., & Kim, J.-S. (2014). Cas-OFFinder: a fast and versatile algorithm that searches for potential off-target sites of Cas9 RNA-guided endonucleases. Bioinformatics, 30(10), 1473–1475. <https://doi.org/10.1093/bioinformatics/btu048>

Barratt, B. (2011). Assessing safety of biological control introductions. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources, 6(042).

<https://doi.org/10.1079/PAVSNNR20116042>

Barratt, B. I. P., Howarth, F. G., Withers, T. M., Kean, J. M., & Ridley, G. S. (2010). Progress in risk assessment for classical biological control. Biological Control, 52(3), 245–254.

<https://doi.org/10.1016/j.biocontrol.2009.02.012>

Barrett, L. G., Legros, M., Kumaran, N., Glassop, D., Raghu, S., & Gardiner, D. M. (2019). Gene drives in plants: opportunities and challenges for weed control and engineered resilience. *Proceedings of the Royal Society B: Biological Sciences*, 286(1911), 20191515. <https://doi.org/10.1098/rspb.2019.1515>

Beaghton, A. K., Hammond, A., Nolan, T., Crisanti, A., & Burt, A. (2019). Gene drive for population genetic control: non-functional resistance and parental effects. *Proceedings of the Royal Society B: Biological Sciences*, 286(1914), 20191586. <https://doi.org/10.1098/rspb.2019.1586>

Beaghton, A., Beaghton, P. J., & Burt, A. (2017). Vector control with driving Y chromosomes: modelling the evolution of resistance. *Malaria Journal*, 16(1), 286. <https://doi.org/10.1186/s12936-017-1932-7>

Benedict, M. (2003). The first releases of transgenic mosquitoes: an argument for the sterile insect technique. *Trends in Parasitology*, 19(8), 349–355. [https://doi.org/10.1016/S1471-4922\(03\)00144-2](https://doi.org/10.1016/S1471-4922(03)00144-2)

Benedict, M., D'Abbs, P., Dobson, S., Gottlieb, M., Harrington, L., Higgs, S., James, A., James, S., Knols, B., Lavery, J., O'Neill, S., Scott, T., Takken, W., & Toure, Y. (2008). Guidance for Contained Field Trials of Vector Mosquitoes Engineered to Contain a Gene Drive System: Recommendations of a Scientific Working Group. *Vector-Borne and Zoonotic Diseases*, 8(2), 127–166.

<https://doi.org/10.1089/vbz.2007.0273>

Benedict, M. Q., Burt, A., Capurro, M. L., De Barro, P., Handler, A. M., Hayes, K. R., Marshall, J. M., Tabachnick, W. J., & Adelman, Z. N. (2018). Recommendations for Laboratory Containment and Management of Gene Drive Systems in Arthropods. *Vector-Borne and Zoonotic Diseases*, 18(1), 2–13. <https://doi.org/10.1089/vbz.2017.2121>

Biémont, C. (2010). A Brief History of the Status of Transposable Elements: From Junk DNA to Major Players in Evolution: Figure 1.—. *Genetics*, 186(4), 1085–1093.

<https://doi.org/10.1534/genetics.110.124180>

Bourguignon, D. (2015). The precautionary principle: Definitions, applications and governance. <https://doi.org/10.2861/821468>

Buchman, A., Marshall, J. M., Ostrovski, D., Yang, T., & Akbari, O. S. (2018). Synthetically engineered Medea gene drive system in the worldwide crop pest *Drosophila suzukii*. *Proceedings of the National Academy of Sciences*, 115(18), 4725–4730. <https://doi.org/10.1073/pnas.1713139115>

Buchthal, J., Evans, S. W., Lunshof, J., Telford, S. R., & Esveld, K. M. (2019). Mice Against Ticks: an experimental community-guided effort to prevent tick-borne disease by altering the shared environment. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 374(1772), 20180105. <https://doi.org/10.1098/rstb.2018.0105>

Bull, J. J. (2015). Evolutionary decay and the prospects for long-term disease intervention using engineered insect vectors: Figure 1. *Evolution, Medicine, and Public Health*, 2015(1), 152–166. <https://doi.org/10.1093/emph/eov013>

Bull, J. J., & Malik, H. S. (2017). The gene drive bubble: New realities. *PLOS Genetics*, 13(7), e1006850. <https://doi.org/10.1371/journal.pgen.1006850>

Bull, J. J., Remien, C. H., Gomulkiewicz, R., & Krone, S. M. (2019). Spatial structure undermines parasite suppression by gene drive cargo. *PeerJ*, 7, e7921. <https://doi.org/10.7717/peerj.7921>

Bull, J. J., Remien, C. H., & Krone, S. M. (2019). Gene-drive-mediated extinction is thwarted by population structure and evolution of sib mating. *Evolution, Medicine, and Public Health*, 2019(1), 66–81. <https://doi.org/10.1093/emph/eoz014>

Burt, A. (2003). Site-specific selfish genes as tools for the control and genetic engineering of natural populations. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 270(1518), 921–928. <https://doi.org/10.1098/rspb.2002.2319>

Burt, A., Coulibaly, M., Crisanti, A., Diabate, A., & Kayondo, J. K. (2018). Gene drive to reduce malaria transmission in sub-Saharan Africa. *Journal of Responsible Innovation*, 5(sup1), S66–S80. <https://doi.org/10.1080/23299460.2017.1419410>

Burt, A., & Crisanti, A. (2018). Gene Drive: Evolved and Synthetic. *ACS Chemical Biology*, 13(2), 343–346. <https://doi.org/10.1021/acscchembio.7b01031>

Campbell, K. J., Beek, J., Eason, C. T., Glen, A. S., Godwin, J., Gould, F., Holmes, N. D., Howald, G. R., Madden, F. M., Ponder, J. B., Threadgill, D. W., Wegmann, A. S., & Baxter, G. S. (2015). The next generation of rodent eradications: Innovative technologies and tools to improve species specificity and increase their feasibility on islands. *Biological Conservation*, 185, 47–58.

<https://doi.org/10.1016/j.biocon.2014.10.016>

Carballar-Lejarazú, R., & James, A. A. (2017). Population modification of Anopheline species to control malaria transmission. *Pathogens and Global Health*, 111(8), 424–435.

<https://doi.org/10.1080/20477724.2018.1427192>

Carter, S. ., & Friedman, R. M. (2016). Policy and Regulatory Issues for Gene Drives in Insects Workshop Report. <https://www.jcvi.org/sites/default/files/assets/projects/policy-and-regulatory-issues-for-gene-drives-in-insects/report-complete.pdf>

Cash, S. A., Lorenzen, M. D., & Gould, F. (2019). The distribution and spread of naturally occurring Medea selfish genetic elements in the United States. *Ecology and Evolution*, 9(24), 14407–14416. <https://doi.org/10.1002/ece3.5876>

Champer, J., Buchman, A., & Akbari, O. S. (2016). Cheating evolution: engineering gene drives to manipulate the fate of wild populations. *Nature Reviews Genetics*, 17(3), 146–159. <https://doi.org/10.1038/nrg.2015.34>

Champer, J., Chung, J., Lee, Y. L., Liu, C., Yang, E., Wen, Z., Clark, A. G., & Messer, P. W. (2019). Molecular safeguarding of CRISPR gene drive experiments. *eLife*, 8. <https://doi.org/10.7554/eLife.41439>

Champer, J., Kim, I., Champer, S., Clark, A., & Messer, P. W. (2019). Suppression gene drive in continuous space can result in unstable persistence of both drive and wild-type alleles. *BioRxiv*, pre-print. <https://doi.org/https://doi.org/10.1101/769810>

Champer, J., Liu, J., Oh, S. Y., Reeves, R., Luthra, A., Oakes, N., Clark, A. G., & Messer, P. W. (2018). Reducing resistance allele formation in CRISPR gene drive. *Proceedings of the National Academy of Sciences*, 115(21), 5522–5527. <https://doi.org/10.1073/pnas.1720354115>

Champer, J., Reeves, R., Oh, S. Y., Liu, C., Liu, J., Clark, A. G., & Messer, P. W. (2017). Novel CRISPR/Cas9 gene drive constructs reveal insights into mechanisms of resistance allele formation and drive efficiency in genetically diverse populations. *PLOS Genetics*, 13(7), e1006796. <https://doi.org/10.1371/journal.pgen.1006796>

Collins, C. M., Bonds, J. A. S., Quinlan, M. M., & Mumford, J. D. (2019). Effects of the removal or reduction in density of the malaria mosquito, *Anopheles gambiae s.l.*, on interacting predators and competitors in local ecosystems. *Medical and Veterinary Entomology*, 33(1), 1–15.

<https://doi.org/10.1111/mve.12327>

Collins, J. P. (2018). Gene drives in our future: challenges of and opportunities for using a self-sustaining technology in pest and vector management. *BMC Proceedings*, 12(S8), 9. <https://doi.org/10.1186/s12919-018-0110-4>

Commission of the European Communities. (2000). Communication from the Commission on the precautionary principle (p. 28). <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:52000DC0001&from=EX>

Cooper, T. F., & Heinemann, J. A. (2005). Selection for plasmid post-segregational killing depends on multiple infection: evidence for the selection of more virulent parasites through parasite-level competition. *Proceedings of the Royal Society B: Biological Sciences*, 272(1561), 403–410.

<https://doi.org/10.1098/rspb.2004.2921>

Cooper, T. F., & Heinemann, J. A. (2000). Postsegregational killing does not increase plasmid stability but acts to mediate the exclusion of competing plasmids. *Proceedings of the National Academy of Sciences*, 97(23), 12643–12648. <https://doi.org/10.1073/pnas.220077897>

Cotter, J., Kawall, K., & Then, C. (2020). New genetic engineering technologies. Report of the results from the RAGES project 2016-2019. http://www.testbiotech.org/projekt_rages

Critical Scientists Switzerland, European Network of Scientists for Social and Environmental Responsibility, & Vereinigung Deutscher Wissenschaftler. (2019). Gene drives: A report on their science, applications, social aspects, ethics and regulations. <https://genedrives.ch/report>

David, A. S., Kaser, J. M., Morey, A. C., Roth, A. M., & Andow, D. A. (2013). Release of genetically engineered insects: a framework to identify potential ecological effects. *Ecology and Evolution*, 3(11), 4000–4015. <https://doi.org/10.1002/ece3.737>

Davis, K. M., Pattanayak, V., Thompson, D. B., Zuris, J. A., & Liu, D. R. (2015). Small molecule-triggered Cas9 protein with improved genome-editing specificity. *Nature Chemical Biology*, 11(5), 316–318. <https://doi.org/10.1038/nchembio.1793>

de Jong, T. J. (2017). Gene drives do not always increase in frequency: from genetic models to risk assessment. *Journal of Consumer Protection and Food Safety*, 12(4), 299–307. <https://doi.org/10.1007/s00003-017-1131-z>

de Lorenzo, V. (2017). Seven microbial bio-processes to help the planet. *Microbial Biotechnology*, 10(5), 995–998. <https://doi.org/10.1111/1751-7915.12816>

Dearden, P. K., Gemmell, N. J., Mercier, O. R., Lester, P. J., Scott, M. J., Newcomb, R. D., Buckley, T. R., Jacobs, J. M. E., Goldson, S. G., & Penman, D. R. (2018). The potential for the use of gene drives for pest control in New Zealand: a perspective. *Journal of the Royal Society of New Zealand*, 48(4), 225–244. <https://doi.org/10.1080/03036758.2017.1385030>

Deredec, A., Godfray, H. C. J., & Burt, A. (2011). Requirements for effective malaria control with homing endonuclease genes. *Proceedings of the National Academy of Sciences*, 108(43), E874–E880.

<https://doi.org/10.1073/pnas.1110717108>

Devos, Y., Craig, W., Devlin, R. H., Ippolito, A., Leggatt, R. A., Romeis, J., Shaw, R., Svendsen, C., & Topping, C. J. (2019). Using problem formulation for fit- for- purpose pre- market environmental risk assessments of regulated stressors. EFSA Journal, 17. <https://doi.org/10.2903/j.efsa.2019.e170708>

Dhole, S., Vella, M. R., Lloyd, A. L., & Gould, F. (2018). Invasion and migration of spatially self-limiting gene drives: A comparative analysis. Evolutionary Applications, 11(5), 794–808.
<https://doi.org/10.1111/eva.12583>

Driesche, R. (2016). Non-target effects of insect biocontrol agents and trends in host specificity since 1985. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources, 11(044). <https://doi.org/10.1079/PAVSNNR201611044>

Drury, D. W., Dapper, A. L., Siniard, D. J., Zentner, G. E., & Wade, M. J. (2017). CRISPR/Cas9 gene drives in genetically variable and nonrandomly mating wild populations. Science Advances, 3(5), e1601910. <https://doi.org/10.1126/sciadv.1601910>

Eckhoff, P. A., Wenger, E. A., Godfray, H. C. J., & Burt, A. (2017). Impact of mosquito gene drive on malaria elimination in a computational model with explicit spatial and temporal dynamics. Proceedings of the National Academy of Sciences, 114(2), E255–E264. <https://doi.org/10.1073/pnas.1611064114>

Edgington, M. P., & Alphey, L. S. (2018). Population dynamics of engineered underdominance and killer-rescue gene drives in the control of disease vectors. PLOS Computational Biology, 14(3), e1006059.
<https://doi.org/10.1371/journal.pcbi.1006059>

EFSA Panel on Genetically Modified Organisms (GMO). (2010). Scientific Opinion on the assessment of potential impacts of genetically modified plants on non-target organisms. EFSA Journal, 8(11), 1877.
<https://doi.org/10.2903/j.efsa.2010.1877>

EFSA Panel on Genetically Modified Organisms (GMO). (2013). Guidance on the environmental risk assessment of genetically modified animals. EFSA Journal, 11(5), 3200.
<https://doi.org/10.2903/j.efsa.2013.3200>

Emerson, C., James, S., Littler, K., & Randazzo, F. (Fil). (2017). Principles for gene drive research. Science, 358(6367), 1135–1136. <https://doi.org/10.1126/science.aap9026>

Esvelt, K. M., & Gemmell, N. J. (2017). Conservation demands safe gene drive. PLOS Biology, 15(11), e2003850. <https://doi.org/10.1371/journal.pbio.2003850>

Esvelt, K. M., Smidler, A. L., Catteruccia, F., & Church, G. M. (2014). Concerning RNA-guided gene drives for the alteration of wild populations. eLife, 3. <https://doi.org/10.7554/eLife.03401>

Farooque, M., Barnhill-Dilling, S. K., Shapiro, J., & Delborne, J. (2019). Exploring Stakeholder Perspectives on the Development of a Gene Drive Mouse for Biodiversity Protection on Islands.
<http://go.ncsu.edu/ges-gene-drive-workshop>

Franz, A. W. E., Sanchez-Vargas, I., Piper, J., Smith, M. R., Khoo, C. C. H., James, A. A., & Olson, K. E. (2009). Stability and loss of a virus resistance phenotype over time in transgenic mosquitoes harbouring an antiviral effector gene. Insect Molecular Biology, 18(5), 661–672. <https://doi.org/10.1111/j.1365-2583.2009.00908.x>

Friedman, R. M., Marshall, J. M., & Akbari, O. S. (2020). Gene Drives: New and Improved. Issues in Science and Technology, XXXVI(2). <https://issues.org/issue/36-2/>

Frieß, J. L., von Gleich, A., & Giese, B. (2019). Gene drives as a new quality in GMO releases—a comparative technology characterization. *PeerJ*, 7, e6793. <https://doi.org/10.7717/peerj.6793>

Galizi, R., Hammond, A., Kyrou, K., Taxiarchi, C., Bernardini, F., O'Loughlin, S. M., Papathanos, P.-A., Nolan, T., Windbichler, N., & Crisanti, A. (2016). A CRISPR-Cas9 sex-ratio distortion system for genetic control. *Scientific Reports*, 6(1), 31139. <https://doi.org/10.1038/srep31139>

Gantz, V. M., & Bier, E. (2015). The mutagenic chain reaction: A method for converting heterozygous to homozygous mutations. *Science*, 348(6233), 442–444. <https://doi.org/10.1126/science.aaa5945>

Gantz, V. M., Jasinskiene, N., Tatarenkova, O., Fazekas, A., Macias, V. M., Bier, E., & James, A. A. (2015). Highly efficient Cas9-mediated gene drive for population modification of the malaria vector mosquito *Anopheles stephensi*. *Proceedings of the National Academy of Sciences*, 112(49), E6736–E6743. <https://doi.org/10.1073/pnas.1521077112>

George, D. R., Kuiken, T., & Delborne, J. A. (2019). Articulating ‘free, prior and informed consent’ (FPIC) for engineered gene drives. *Proceedings of the Royal Society B: Biological Sciences*, 286(1917), 20191484. <https://doi.org/10.1098/rspb.2019.1484>

Godfray, H. C. J., North, A., & Burt, A. (2017). How driving endonuclease genes can be used to combat pests and disease vectors. *BMC Biology*, 15(1), 81. <https://doi.org/10.1186/s12915-017-0420-4>

Godwin, J., Serr, M., Barnhill-Dilling, S. K., Blondel, D. V., Brown, P. R., Campbell, K., Delborne, J., Lloyd, A. L., Oh, K. P., Prowse, T. A. A., Saah, R., & Thomas, P. (2019). Rodent gene drives for conservation: opportunities and data needs. *Proceedings of the Royal Society B: Biological Sciences*, 286(1914), 20191606. <https://doi.org/10.1098/rspb.2019.1606>

Grunwald, H. A., Gantz, V. M., Poplawski, G., Xu, X.-R. S., Bier, E., & Cooper, K. L. (2019). Super-Mendelian inheritance mediated by CRISPR–Cas9 in the female mouse germline. *Nature*, 566(7742), 105–109. <https://doi.org/10.1038/s41586-019-0875-2>

Gutzmann, N., Elsensohn, J. E., Barnes, J. C., Baltzegar, J., Jones, M. S., & Sudweeks, J. (2017). CRISPR- based gene drive in agriculture will face technical and governance challenges. *EMBO Reports*, 18(9), 1479–1480. <https://doi.org/10.15252/embr.201744661>

Hammond, A. M., Kyrou, K., Bruttini, M., North, A., Galizi, R., Karlsson, X., Kranjc, N., Carpi, F. M., D'Aurizio, R., Crisanti, A., & Nolan, T. (2017). The creation and selection of mutations resistant to a gene drive over multiple generations in the malaria mosquito. *PLOS Genetics*, 13(10), e1007039. <https://doi.org/10.1371/journal.pgen.1007039>

Hammond, A., Galizi, R., Kyrou, K., Simoni, A., Siniscalchi, C., Katsanos, D., Gribble, M., Baker, D., Marois, E., Russell, S., Burt, A., Windbichler, N., Crisanti, A., & Nolan, T. (2016). A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*. *Nature Biotechnology*, 34(1), 78–83. <https://doi.org/10.1038/nbt.3439>

Hartley, S., Thizy, D., Ledingham, K., Coulibaly, M., Diabaté, A., Dicko, B., Diop, S., Kayondo, J., Namukwaya, A., Nourou, B., & Paré Toé, L. (2019). Knowledge engagement in gene drive research for malaria control. *PLOS Neglected Tropical Diseases*, 13(4), e0007233. <https://doi.org/10.1371/journal.pntd.0007233>

Harvey- Samuel, T., Norman, V. C., Carter, R., Lovett, E., & Alphey, L. (2019). Identification and characterization of a *Masculinizer* homologue in the diamondback moth, *Plutella xylostella*. *Insect Molecular Biology*, imb.12628. <https://doi.org/10.1111/imb.12628>

Harvey-Samuel, T. O., Campbell, K. J., Edgington, M., & Alphey, L. (2019). Trialling gene drives to control invasive species: what, where and how? In C. R. Veitch, M. N. Clout, A. R. Martin, J. C. Russell, & C. J. West (Eds.), Island invasives: scaling up to meet the challenge (Occasional, pp. 618–627). International Union for Conservation of Nature.

https://digitalcommons.unl.edu/cgi/viewcontent.cgi?article=3234&context=icwdm_usdanwrc

Haut Conseil des Biotechnologies. (2017). Scientific Opinion in response to the referral of 12 October 2015 concerning the use of genetically modified mosquitoes for vector control (p. 142).

<http://www.hautconseildesbiotechnologies.fr/en/avis/avis-relatif-a-lutilisation-moustiques-gm-dans-cadre-lutte-antivectorielle>

Hayes, K. R., Hosack, G. R., Dana, G. V., Foster, S. D., Ford, J. H., Thresher, R., Ickowicz, A., Peel, D., Tizard, M., De Barro, P., Strive, T., & Dambacher, J. M. (2018). Identifying and detecting potentially adverse ecological outcomes associated with the release of gene-drive modified organisms. *Journal of Responsible Innovation*, 5(sup1), S139–S158. <https://doi.org/10.1080/23299460.2017.1415585>

Heffel, M. G., & Finnigan, G. C. (2019). Mathematical modeling of self-contained CRISPR gene drive reversal systems. *Scientific Reports*, 9(1), 20050. <https://doi.org/10.1038/s41598-019-54805-8>

Heinemann, J. A. (2019). Should dsRNA treatments applied in outdoor environments be regulated? *Environment International*, 132, 104856. <https://doi.org/10.1016/j.envint.2019.05.050>

Heinemann, J. A. (2007). A typology of the effects of (trans)gene flow on the conservation and sustainable use of genetic resources (No. 35; Background Study Paper). <http://www.fao.org/3/a-k0153e.pdf>

Heinemann, J. A., & Walker, S. (2019). Environmentally applied nucleic acids and proteins for purposes of engineering changes to genes and other genetic material. *Biosafety and Health*. <https://doi.org/10.1016/j.bsheal.2019.09.003>

Hoffmann, A. A., Ankeny, R., Edwards, O., Frommer, M., Hayes, K. R., Higgins, T., Mayo, O., Meek, S., Robin, C., Sheppard, A., & Small, I. (2017). Synthetic gene drives in Australia: implications of emerging technologies. <https://www.science.org.au/support/analysis/reports/synthetic-gene-drives-australia-implications-emerging-technologies>

Houck, M., Clark, J., Peterson, K., & Kidwell, M. (1991). Possible horizontal transfer of *Drosophila* genes by the mite *Proctolaelaps regalis*. *Science*, 253(5024), 1125–1128. <https://doi.org/10.1126/science.1653453>

Huang, Y., Magori, K., Lloyd, A. L., & Gould, F. (2007). Introducing desirable transgenes into insect populations using Y-linked meiotic drive> A theoretical assessment. *Evolution*, 61(4), 717–726. <https://doi.org/10.1111/j.1558-5646.2007.00075.x>

Hudson, M., Mead, A. T. P., Chagné, D., Roskruge, N., Morrison, S., Wilcox, P. L., & Allan, A. C. (2019). Indigenous Perspectives and Gene Editing in Aotearoa New Zealand. *Frontiers in Bioengineering and Biotechnology*, 7. <https://doi.org/10.3389/fbioe.2019.00070>

Hurst, L. D. (2019). A century of bias in genetics and evolution. *Heredity*, 123(1), 33–43. <https://doi.org/10.1038/s41437-019-0194-2>

International Finance Corporation of the World Bank Group. (2012). A guide to biodiversity for the private sector (p. 3). <https://www.ifc.org/wps/wcm/connect/9608497e-56e8-4074-bab6-45c61a36a4ad/ESIA.pdf?MOD=AJPERES&CVID=jkCYZ3G>

James, A. A. (2005). Gene drive systems in mosquitoes: rules of the road. *Trends in Parasitology*, 21(2), 64–67. <https://doi.org/10.1016/j.pt.2004.11.004>

James, S., Collins, F. H., Welkhoff, P. A., Emerson, C., Godfray, H. C. J., Gottlieb, M., Greenwood, B., Lindsay, S. W., Mbogo, C. M., Okumu, F. O., Quemada, H., Savadogo, M., Singh, J. A., Tountas, K. H., & Touré, Y. T. (2018). Pathway to Deployment of Gene Drive Mosquitoes as a Potential Biocontrol Tool for Elimination of Malaria in Sub-Saharan Africa: Recommendations of a Scientific Working Group †. *The American Journal of Tropical Medicine and Hygiene*, 98(6_Suppl), 1–49. <https://doi.org/10.4269/ajtmh.18-0083>

James, S., & Tountas, K. (2018). Using Gene Drive Technologies to Control Vector-Borne Infectious Diseases. *Sustainability*, 10(12), 4789. <https://doi.org/10.3390/su10124789>

Jones, M. S., Delborne, J. A., Elsensohn, J., Mitchell, P. D., & Brown, Z. S. (2019). Does the U.S. public support using gene drives in agriculture? And what do they want to know? *Science Advances*, 5(9), eaau8462. <https://doi.org/10.1126/sciadv.aau8462>

Kandul, N. P., Liu, J., Buchman, A., Gantz, V. M., Bier, E., & Akbari, O. S. (2019). Assessment of a Split Homing Based Gene Drive for Efficient Knockout of Multiple Genes. *G3: Genes|Genomes|Genetics*, g3.400985.2019. <https://doi.org/10.1534/g3.119.400985>

Kolopack, P. A., & Lavery, J. V. (2017). Informed consent in field trials of gene-drive mosquitoes. *Gates Open Research*, 1, 14. <https://doi.org/10.12688/gatesopenres.12771.1>

Koo, T., Lee, J., & Kim, J.-S. (2015). Measuring and Reducing Off-Target Activities of Programmable Nucleases Including CRISPR-Cas9. *Molecules and Cells*, 38(6), 475–481. <https://doi.org/10.14348/molcells.2015.0103>

Kopf, R. K., Nimmo, D. G., Humphries, P., Baumgartner, L. J., Bode, M., Bond, N. R., Byrom, A. E., Cucherousset, J., Keller, R. P., King, A. J., McGinness, H. M., Moyle, P. B., & Olden, J. D. (2017). Confronting the risks of large-scale invasive species control. *Nature Ecology & Evolution*, 1(6), 0172. <https://doi.org/10.1038/s41559-017-0172>

Kuzma, J., Gould, F., Brown, Z., Collins, J., Delborne, J., Frow, E., Esvelt, K., Guston, D., Leitschuh, C., Oye, K., & Stauffer, S. (2018). A roadmap for gene drives: using institutional analysis and development to frame research needs and governance in a systems context. *Journal of Responsible Innovation*, 5(sup1), S13–S39. <https://doi.org/10.1080/23299460.2017.1410344>

Kuzma, J. (2019). Procedurally Robust Risk Assessment Framework for Novel Genetically Engineered Organisms and Gene Drives. *Regulation & Governance*, rego.12245. <https://doi.org/10.1111/rego.12245>

Kyrou, K., Hammond, A. M., Galizi, R., Kranjc, N., Burt, A., Beaghton, A. K., Nolan, T., & Crisanti, A. (2018). A CRISPR–Cas9 gene drive targeting doublesex causes complete population suppression in caged *Anopheles gambiae* mosquitoes. *Nature Biotechnology*, 36(11), 1062–1066. <https://doi.org/10.1038/nbt.4245>

Leftwich, P. T., Edgington, M. P., Harvey-Samuel, T., Carabajal Paladino, L. Z., Norman, V. C., & Alphey, L. (2018). Recent advances in threshold-dependent gene drives for mosquitoes. *Biochemical Society Transactions*, 46(5), 1203–1212. <https://doi.org/10.1042/BST20180076>

Legros, M., Xu, C., Morrison, A., Scott, T. W., Lloyd, A. L., & Gould, F. (2013). Modeling the Dynamics of a Non-Limited and a Self-Limited Gene Drive System in Structured *Aedes aegypti* Populations. *PLoS ONE*, 8(12), e83354. <https://doi.org/10.1371/journal.pone.0083354>

Li, M., Yang, T., Kandul, N. P., Bui, M., Gamez, S., Raban, R., Bennett, J., Sánchez C, H. M., Lanzaro, G. C., Schmidt, H., Lee, Y., Marshall, J. M., & Akbari, O. S. (2020). Development of a confinable gene drive system in the human disease vector *Aedes aegypti*. *ELife*, 9. <https://doi.org/10.7554/eLife.51701>

Lindholm, A. K., Dyer, K. A., Firman, R. C., Fishman, L., Forstmeier, W., Holman, L., Johannesson, H., Knief, U., Kokko, H., Larracuente, A. M., Manser, A., Montchamp-Moreau, C., Petrosyan, V. G., Pomiankowski, A., Presgraves, D. C., Safranova, L. D., Sutter, A., Unckless, R. L., Verspoor, R. L., ... Price, T. A. R. (2016). The Ecology and Evolutionary Dynamics of Meiotic Drive. *Trends in Ecology & Evolution*, 31(4), 315–326. <https://doi.org/10.1016/j.tree.2016.02.001>

Liu, Y., & Stewart, C. N. (2019). An exposure pathway- based risk assessment system for GM plants. *Plant Biotechnology Journal*, 17(10), 1859–1861. <https://doi.org/10.1111/pbi.13146>

Louda, S. M., Pemberton, R. W., Johnson, M. T., & Follett, P. A. (2003). Nontarget effects – The Achilles' heel of biological control? Retrospective Analyses to Reduce Risk Associated with Biocontrol Introductions. *Annual Review of Entomology*, 48(1), 365–396.
<https://doi.org/10.1146/annurev.ento.48.060402.102800>

Ma, E., Harrington, L. B., O'Connell, M. R., Zhou, K., & Doudna, J. A. (2015). Single-Stranded DNA Cleavage by Divergent CRISPR-Cas9 Enzymes. *Molecular Cell*, 60(3), 398–407.
<https://doi.org/10.1016/j.molcel.2015.10.030>

Macias, V. M., Jimenez, A. J., Burini-Kojin, B., Pledger, D., Jasinskiene, N., Phong, C. H., Chu, K., Fazekas, A., Martin, K., Marinotti, O., & James, A. A. (2017). nanos-Driven expression of piggyBac transposase induces mobilization of a synthetic autonomous transposon in the malaria vector mosquito, *Anopheles stephensi*. *Insect Biochemistry and Molecular Biology*, 87, 81–89.
<https://doi.org/10.1016/j.ibmb.2017.06.014>

Macias, V., Ohm, J., & Rasgon, J. (2017). Gene Drive for Mosquito Control: Where Did It Come from and Where Are We Headed? *International Journal of Environmental Research and Public Health*, 14(9), 1006.
<https://doi.org/10.3390/ijerph14091006>

Manser, A., Cornell, S. J., Sutter, A., Blondel, D. V., Serr, M., Godwin, J., & Price, T. A. R. (2019). Controlling invasive rodents via synthetic gene drive and the role of polyandry. *Proceedings of the Royal Society B: Biological Sciences*, 286(1909), 20190852. <https://doi.org/10.1098/rspb.2019.0852>

Marshall, J. M., & Hay, B. A. (2011). Inverse Medea as a Novel Gene Drive System for Local Population Replacement: A Theoretical Analysis. *Journal of Heredity*, 102(3), 336–341.
<https://doi.org/10.1093/jhered/esr019>

Marshall, J. M. (2009). The effect of gene drive on containment of transgenic mosquitoes. *Journal of Theoretical Biology*, 258(2), 250–265. <https://doi.org/10.1016/j.jtbi.2009.01.031>

Marshall, J. M., & Akbari, O. S. (2018). Can CRISPR-Based Gene Drive Be Confined in the Wild? A Question for Molecular and Population Biology. *ACS Chemical Biology*, 13(2), 424–430.
<https://doi.org/10.1021/acscchembio.7b00923>

Marshall, J. M., Buchman, A., Sánchez C, H. M., & Akbari, O. S. (2017). Overcoming evolved resistance to population-suppressing homing-based gene drives. *Scientific Reports*, 7(1), 3776.
<https://doi.org/10.1038/s41598-017-02744-7>

Marshall, J. M., & Hay, B. A. (2012). Confinement of gene drive systems to local populations: A comparative analysis. *Journal of Theoretical Biology*, 294, 153–171.

<https://doi.org/10.1016/j.jtbi.2011.10.032>

Mathematical Ecology Group. (n.d.). Ecological Risks of Gene Drive Technologies.

https://merg.zoo.ox.ac.uk/sites/default/files/GeneDrivePolicyBrief_PRINT.pdf

Meghani, Z. (2019). Autonomy of Nations and Indigenous Peoples and the Environmental Release of Genetically Engineered Animals with Gene Drives. *Global Policy*, 10(4), 554–568.

<https://doi.org/10.1111/1758-5899.12699>

Min, J., Noble, C., Najjar, D., & Esveld, K. M. (2017). Daisy quorum drives for the genetic restoration of wild populations. *BioRxiv*, pre-print. <https://doi.org/doi.org/10.1101/115618>

Min, J., Smidler, A. L., Najjar, D., & Esveld, K. M. (2018). Harnessing gene drive. *Journal of Responsible Innovation*, 5(sup1), S40–S65. <https://doi.org/10.1080/23299460.2017.1415586>

Moro, D., Byrne, M., Kennedy, M., Campbell, S., & Tizard, M. (2018). Identifying knowledge gaps for gene drive research to control invasive animal species: The next CRISPR step. *Global Ecology and Conservation*, 13, e00363. <https://doi.org/10.1016/j.gecco.2017.e00363>

Murphy, B., Jensen, C., Murray, J., & De Barro, P. (2010). Risk Analysis on the Australian release of *Aedes aegypti* (L.) (Diptera: Culicidae) containing *Wolbachia*.

<http://www.eliminatedengue.com/library/publication/document/riskanalysisfinalreportcsiro.pdf>

Nash, A., Urdaneta, G. M., Beaghton, A. K., Hoermann, A., Papathanos, P. A., Christophides, G. K., & Windbichler, N. (2019). Integral gene drives for population replacement. *Biology Open*, 8(1), bio037762. <https://doi.org/10.1242/bio.037762>

National Academies of Sciences Engineering and Medicine. (2016). Gene Drives on the Horizon: Advancing Science, Navigating Uncertainty, and Aligning Research with Public Values. National Academies Press. <https://doi.org/10.17226/23405>

National Research Council. (1983). Risk Assessment in the Federal Government. National Academies Press. <https://doi.org/10.17226/366>

National Research Council. (1996). Understanding Risk. National Academies Press. <https://doi.org/10.17226/5138>

Neve, P. (2018). Gene drive systems: do they have a place in agricultural weed management? *Pest Management Science*, 74(12), 2671–2679. <https://doi.org/10.1002/ps.5137>

Nihongaki, Y., Otabe, T., & Sato, M. (2018). Emerging Approaches for Spatiotemporal Control of Targeted Genome with Inducible CRISPR-Cas9. *Analytical Chemistry*, 90(1), 429–439.

<https://doi.org/10.1021/acs.analchem.7b04757>

Noble, C., Adlam, B., Church, G. M., Esveld, K. M., & Nowak, M. A. (2018). Current CRISPR gene drive systems are likely to be highly invasive in wild populations. *ELife*, 7. <https://doi.org/10.7554/eLife.33423>

Noble, C., Min, J., Olejarz, J., Buchthal, J., Chavez, A., Smidler, A. L., DeBenedictis, E. A., Church, G. M., Nowak, M. A., & Esveld, K. M. (2019). Daisy-chain gene drives for the alteration of local populations. *Proceedings of the National Academy of Sciences*, 116(17), 8275–8282.

<https://doi.org/10.1073/pnas.1716358116>

Noble, C., Olejarz, J., Esvelt, K. M., Church, G. M., & Nowak, M. A. (2017). Evolutionary dynamics of CRISPR gene drives. *Science Advances*, 3(4), e1601964. <https://doi.org/10.1126/sciadv.1601964>

North, A. R., Burt, A., & Godfray, H. C. J. (2019). Modelling the potential of genetic control of malaria mosquitoes at national scale. *BMC Biology*, 17(1), 26. <https://doi.org/10.1186/s12915-019-0645-5>

North, A., Burt, A., & Godfray, H. C. J. (2013). Modelling the spatial spread of a homing endonuclease gene in a mosquito population. *Journal of Applied Ecology*, n/a-n/a. <https://doi.org/10.1111/1365-2664.12133>

Oberhofer, G., Ivy, T., & Hay, B. A. (2019). Cleave and Rescue, a novel selfish genetic element and general strategy for gene drive. *Proceedings of the National Academy of Sciences*, 116(13), 6250–6259. <https://doi.org/10.1073/pnas.1816928116>

OECD. (2018). Safety Assessment of Transgenic Organisms in the Environment, Volume 8 OECD Consensus Document of the Biology of Mosquito *Aedes aegypti*. OECD.
<https://doi.org/10.1787/9789264302235-en>

O'Neill, S. L., Ryan, P. A., Turley, A. P., Wilson, G., Retzki, K., Iturbe-Ormaetxe, I., Dong, Y., Kenny, N., Paton, C. J., Ritchie, S. A., Brown-Kenyon, J., Stanford, D., Wittmeier, N., Jewell, N. P., Tanamas, S. K., Anders, K. L., & Simmons, C. P. (2019). Scaled deployment of *Wolbachia* to protect the community from dengue and other *Aedes* transmitted arboviruses. *Gates Open Research*, 2, 36. <https://doi.org/10.12688/gatesopenres.12844.3>

Oye, K. A., Esvelt, K., Appleton, E., Catteruccia, F., Church, G., Kuiken, T., Lightfoot, S. B.-Y., McNamara, J., Smidler, A., & Collins, J. P. (2014). Regulating gene drives. *Science*, 345(6197), 626–628. <https://doi.org/10.1126/science.1254287>

Paulo, D. F., Williamson, M. E., Arp, A. P., Li, F., Sagel, A., Skoda, S. R., Sanchez-Gallego, J., Vasquez, M., Quintero, G., Pérez de León, A. A., Belikoff, E. J., Azeredo-Espin, A. M. L., McMillan, W. O., Concha, C., & Scott, M. J. (2019). Specific Gene Disruption in the Major Livestock Pests *Cochliomyia hominivorax* and *Lucilia cuprina* Using CRISPR/Cas9. *G3: Genes|Genomes|Genetics*, 9(9), 3045–3055. <https://doi.org/10.1534/g3.119.400544>

Pham, T. B., Phong, C. H., Bennett, J. B., Hwang, K., Jasinskiene, N., Parker, K., Stillinger, D., Marshall, J. M., Carballar-Lejarazú, R., & James, A. A. (2019). Experimental population modification of the malaria vector mosquito, *Anopheles stephensi*. *PLOS Genetics*, 15(12), e1008440. <https://doi.org/10.1371/journal.pgen.1008440>

Phuc, H., Andreasen, M. H., Burton, R. S., Vass, C., Epton, M. J., Pape, G., Fu, G., Condon, K. C., Scaife, S., Donnelly, C. A., Coleman, P. G., White-Cooper, H., & Alphey, L. (2007). Late-acting dominant lethal genetic systems and mosquito control. *BMC Biology*, 5(1), 11. <https://doi.org/10.1186/1741-7007-5-11>

Price, T. A. R., Verspoor, R., & Wedell, N. (2019). Ancient gene drives: an evolutionary paradox. *Proceedings of the Royal Society B: Biological Sciences*, 286(1917), 20192267. <https://doi.org/10.1098/rspb.2019.2267>

Prowse, T. A. A., Cassey, P., Ross, J. V., Pfitzner, C., Wittmann, T. A., & Thomas, P. (2017). Dodging silver bullets: good CRISPR gene-drive design is critical for eradicating exotic vertebrates. *Proceedings of the Royal Society B: Biological Sciences*, 284(1860), 20170799. <https://doi.org/10.1098/rspb.2017.0799>

Prowse, T. A., Adikusuma, F., Cassey, P., Thomas, P., & Ross, J. V. (2019). A Y-chromosome shredding gene drive for controlling pest vertebrate populations. *eLife*, 8. <https://doi.org/10.7554/eLife.41873>

Quang, T., UyenNinh, T., Van Tuat, N., Viet Hung, N., & Dinh Cuong, N. (2011). Risk Assessment of the Pilot Release of *Aedes aegypti* mosquitoes containing *Wolbachia*.

http://www.eliminatedengue.com/library/publication/document/july_2011_ra_report_eng.pdf

Raybould, A. (2006). Problem formulation and hypothesis testing for environmental risk assessments of genetically modified crops. Environmental Biosafety Research, 5(3), 119–125.

<https://doi.org/10.1051/ebr:2007004>

Redford, K. H., Brooks, T. M., Macfarlane, N. B. W., & Adams, J. S. (Eds.). (2019). Genetic frontiers for conservation: an assessment of synthetic biology and biodiversity conservation: technical assessment. IUCN, International Union for Conservation of Nature. <https://doi.org/10.2305/IUCN.CH.2019.05.en>

Roberts, A., Andrade, P. P. de, Okumu, F., Quemada, H., Savadogo, M., Singh, J. A., & James, S. (2017). Results from the Workshop “Problem Formulation for the Use of Gene Drive in Mosquitoes.” The American Journal of Tropical Medicine and Hygiene, 96(3), 530–533. <https://doi.org/10.4269/ajtmh.16-0726>

Rode, N. O., Estoup, A., Bourguet, D., Courtier-Orgogozo, V., & Débarre, F. (2019). Population management using gene drive: molecular design, models of spread dynamics and assessment of ecological risks. Conservation Genetics, 20(4), 671–690. <https://doi.org/10.1007/s10592-019-01165-5>

Rüdelsheim, P. L. J., & Smets, G. (2018). Gene Drives Experience with gene drive systems that may inform an environmental risk assessment (COGEM Report CGM 2018-03).

<https://cogem.net/en/publication/gene-drives-experience-with-gene-drive-systems-that-may-inform-an-environmental-risk-assessment/>

Sandler, L., & Novitski, E. (1957). Meiotic Drive as an Evolutionary Force. The American Naturalist, 91(857), 105–110. <https://doi.org/10.1086/281969>

Scott, M. J., Gould, F., Lorenzen, M., Grubbs, N., Edwards, O., & O’Brochta, D. (2018). Agricultural production: assessment of the potential use of Cas9-mediated gene drive systems for agricultural pest control. Journal of Responsible Innovation, 5(sup1), S98–S120.

<https://doi.org/10.1080/23299460.2017.1410343>

Secretariat of the Convention on Biological Diversity. (2016). Guidance on Risk Assessment of Living modified Organisms (UNEP/CBD/BS/COP-MOP/8/8/Add.1) (p. 112).

<https://www.cbd.int/doc/meetings/bs/mop-08/official/bs-mop-08-08-add1-en.pdf>

Sim, S. B., Kauwe, A. N., Ruano, R. E. Y., Rendon, P., & Geib, S. M. (2019). The ABCs of CRISPR in Tephritidae: developing methods for inducing heritable mutations in the genera *Anastrepha*, *Bactrocera* and *Ceratitis*. Insect Molecular Biology, 28(2), 277–289. <https://doi.org/10.1111/imb.12550>

Simon, S., Otto, M., & Engelhard, M. (2018). Synthetic gene drive: between continuity and novelty. EMBO Reports, 19(5). <https://doi.org/10.15252/embr.201845760>

Singh, J. A. (2019). Informed consent and community engagement in open field research: lessons for gene drive science. BMC Medical Ethics, 20(1), 54. <https://doi.org/10.1186/s12910-019-0389-3>

Sinkins, S. P., & Gould, F. (2006). Gene drive systems for insect disease vectors. Nature Reviews Genetics, 7(6), 427–435. <https://doi.org/10.1038/nrg1870>

Stroehlein, A. J., Korhonen, P. K., Chong, T. M., Lim, Y. L., Chan, K. G., Webster, B., Rollinson, D., Brindley, P. J., Gasser, R. B., & Young, N. D. (2019). High-quality *Schistosoma haematobium* genome

achieved by single-molecule and long-range sequencing. *GigaScience*, 8(9).

<https://doi.org/10.1093/gigascience/giz108>

Sudweeks, J., Hollingsworth, B., Blondel, D. V., Campbell, K. J., Dhole, S., Eisemann, J. D., Edwards, O., Godwin, J., Howald, G. R., Oh, K. P., Piaggio, A. J., Prowse, T. A. A., Ross, J. V., Saah, J. R., Shiels, A. B., Thomas, P. Q., Threadgill, D. W., Vella, M. R., Gould, F., & Lloyd, A. L. (2019). Locally Fixed Alleles: A method to localize gene drive to island populations. *Scientific Reports*, 9(1), 15821.

<https://doi.org/10.1038/s41598-019-51994-0>

Tanaka, T., Tanaka, N., Nagano, Y., Kanuka, H., Yamamoto, D. S., Yamamoto, N., Nanba, E., & Nishiuchi, T. (2019). Efforts to enhance safety measures for CRISPR/Cas-based gene drive technology in Japan. *Journal of Environment and Safety, advpub*. <https://doi.org/10.11162/daikankyo.E19SC0801>

Teem, J. L., Ambali, A., Glover, B., Ouedraogo, J., Makinde, D., & Roberts, A. (2019). Problem formulation for gene drive mosquitoes designed to reduce malaria transmission in Africa: results from four regional consultations 2016–2018. *Malaria Journal*, 18(1), 347. <https://doi.org/10.1186/s12936-019-2978-5>

Unckless, R. L., Clark, A. G., & Messer, P. W. (2017). Evolution of Resistance Against CRISPR/Cas9 Gene Drive. *Genetics*, 205(2), 827–841. <https://doi.org/10.1534/genetics.116.197285>

US Environmental Protection Agency. (1998). Guidelines for Ecological Risk Assessment (EPA/630/R-95/002F). https://www.epa.gov/sites/production/files/2014-11/documents/eco_risk_assessment1998.pdf

van der Vlugt, C. J. B., Brown, D. D., Lehmann, K., Leunda, A., & Willemarck, N. (2018). A Framework for the Risk Assessment and Management of Gene Drive Technology in Contained Use. *Applied Biosafety*, 23(1), 25–31. <https://doi.org/10.1177/1535676018755117>

Walker, W. E., Harremoës, P., Rotmans, J., van der Sluijs, J. P., van Asselt, M. B. A., Janssen, P., & Krämer von Krauss, M. P. (2003). Defining Uncertainty: A Conceptual Basis for Uncertainty Management in Model-Based Decision Support. *Integrated Assessment*, 4(1), 5–17.

<https://doi.org/10.1076/iaij.4.1.5.16466>

Webster, S. H., Vella, M. R., & Scott, M. J. (2019). No Development and testing of a novel Killer-Rescue self-limiting gene drive system in *Drosophila melanogaster*. *BioRxiv*, Pre-print.

<https://doi.org/https://doi.org/10.1101/680629>

Wedell, N., Price, T. A. R., & Lindholm, A. K. (2019). Gene drive: progress and prospects. *Proceedings of the Royal Society B: Biological Sciences*, 286(1917), 20192709. <https://doi.org/10.1098/rspb.2019.2709>

Werren, J. H. (2011). Selfish genetic elements, genetic conflict, and evolutionary innovation. *Proceedings of the National Academy of Sciences*, 108(Supplement_2), 10863–10870.

<https://doi.org/10.1073/pnas.1102343108>

Westra, J., van der Vlugt, C. J. B., Roesink, C. H., Hogervorst, P. A. M., & Glandorf, D. C. M. (2016). Gene drives: Policy report (RIVM Letter report 2016-0023).

<https://www.rivm.nl/bibliotheek/rapporten/2016-0023.pdf>

Windbichler, N., Menichelli, M., Papathanos, P. A., Thyme, S. B., Li, H., Ulge, U. Y., Hovde, B. T., Baker, D., Monnat, R. J., Burt, A., & Crisanti, A. (2011). A synthetic homing endonuclease-based gene drive system in the human malaria mosquito. *Nature*, 473(7346), 212–215.

<https://doi.org/10.1038/nature09937>

Wolt, J. D., Keese, P., Raybould, A., Fitzpatrick, J. W., Burachik, M., Gray, A., Olin, S. S., Schiemann, J., Sears, M., & Wu, F. (2010). Problem formulation in the environmental risk assessment for genetically modified plants. *Transgenic Research*, 19(3), 425–436. <https://doi.org/10.1007/s11248-009-9321-9>

World Health Organization, Foundation for the National Institutes of Health, & UNICEF/UNDP/World Bank/WHO Special Programme for Research and Training in Tropical Diseases. (2014). The Guidance Framework for testing genetically modified mosquitoes (p. 159).

<https://www.who.int/tdr/publications/year/2014/guide-fmrk-gm-mosquit/en/>

Zentrale Kommission für die Biologische Sicherheit. (2016). Position statement of the ZKBS on the classification of genetic engineering operations for the production and use of higher organisms using recombinant gene drive systems. <http://bch.cbd.int/database/attachment/?id=16811>

Zhao, S., Xing, Z., Liu, Z., Liu, Y., Liu, X., Chen, Z., Li, J., & Yan, R. (2019). Efficient somatic and germline genome engineering of *Bactrocera dorsalis* by the CRISPR/Cas9 system. *Pest Management Science*, 75(7), 1921–1932. <https://doi.org/10.1002/ps.5305>

Zheng, X., Zhang, D., Li, Y., Yang, C., Wu, Y., Liang, X., Liang, Y., Pan, X., Hu, L., Sun, Q., Wang, X., Wei, Y., Zhu, J., Qian, W., Yan, Z., Parker, A. G., Gilles, J. R. L., Bourtzis, K., Bouyer, J., ... Xi, Z. (2019). Incompatible and sterile insect techniques combined eliminate mosquitoes. *Nature*, 572(7767), 56–61. <https://doi.org/10.1038/s41586-019-1407-9>

Online resources

BioTip pilot study: Genetic innovations as a trigger for phase transitions in the population dynamics of animals and plants (GeneTip): <https://www.genetip.de/en/biotip-pilot-study/>

Data61 Ecological and Environmental Risk Assessment Team: <https://data61.csiro.au/en/Our-Research/Programs-and-Facilities/Analytics-and-decision-sciences/DEERA>

European Food Safety Authority Genetically modified organism applications: regulations and guidance: <http://www.efsa.europa.eu/en/applications/gmo/regulationsandguidance>

European Food Safety Authority Public consultations planner:
<https://www.efsa.europa.eu/en/consultations/consultationsplanner>

Foundation for the National Institutes of Health (FNIH) Gene drive researchers forum:
<https://fnih.org/what-we-do/current-lectures-awards-and-events/gene-drive-research-forum>

International Centre for Genetic Engineering and Biotechnology eLearning Showcase: <https://showcase-icgeb.elearning.it/>

International Institute for International Development - Experimental lakes Area: <https://www.iisd.org/ela/>

International Life Sciences Institute (ILSI) Research Foundation “Application of Problem Formulation to the Environmental Risk Assessment of Genetically Engineered Crops” course: <https://ilsirf.org/elearning-courses/pfera/>

National Public Radio – Goats and Soda - VIDEO: The 7 Dwarfs Whistle While They Work To Fight Malaria: <https://www.npr.org/sections/goatsandsoda/2018/08/19/638948839/video-the-7-dwarfs-whistle-while-they-work-to-fight-malaria>

Secretariat of the Convention on Biological Diversity “Guidance on Risk Assessment”:
http://bch.cbd.int/cpb_art15/training.shtml

Secretariat of the Convention on Biological Diversity Submission of information on risk assessment and risk management (2017-2018): https://bch.cbd.int/protocol/cpb_art15_submissions/

Secretariat of the Convention on Biological Diversity Submission of information on risk assessment and risk management (2019): <https://bch.cbd.int/onlineconferences/submissions.shtml>

Target Malaria – Resources: <https://targetmalaria.org/resources/>

TestBiotech RAGES reports: <https://www.testbiotech.org/en/content/research-project-rages>

World Health organization Overview of intervention classes and prototypes/products under Vector Control Advisory Group (VCAG) review for assessment of public health value (as of 1 November 2019):
<https://apps.who.int/iris/bitstream/handle/10665/274451/WHO-CDS-VCAG-2018.03-eng.pdf?ua=1>

World Health organization Vector Control Advisory Group: <https://www.who.int/vector-control/vcag/en/>

Topic 2: Living modified fish

Publications, reports and communications

AquaBounty Technologies Inc. (2018). AquAdvantage® Salmon Environmental Assessment: Supplement to NADA 141-454 to allow the grow-out of AquAdvantage (p. 110).
<https://www.fda.gov/media/112655/download>

Cotter, J., Kawall, K., & Then, C. (2020). New genetic engineering technologies. Report of the results from the RAGES project 2016-2019. http://www.testbiotech.org/projekt_rages

Department of Fisheries and Oceans Canada. (2018). Environmental and Indirect Human Health Risk Assessment of the GloFish® Tetras (*Gymnocorymbus ternetzi*): Five Lines of Transgenic Ornamental Fish. <https://waves-vagues.dfo-mpo.gc.ca/Library/40712928.pdf>

Department of Fisheries and Oceans Canada. (2019). Environmental and Indirect Human Health Risk Assessments for the Manufacture and Grow-out of EO-1 α Salmon, including the AquAdvantage® Salmon, at a Land-Based and Contained Facility near Rollo Bay. <https://waves-vagues.dfo-mpo.gc.ca/Library/4078292x.pdf>

Devlin, R. H., Yesaki, T. Y., Biagi, C. A., Donaldson, E. M., Swanson, P., & Chan, W.-K. (1994). Extraordinary salmon growth. *Nature*, 371(6494), 209–210. <https://doi.org/10.1038/371209a0>

EFSA Panel on Genetically Modified Organisms (GMO). (2013). Guidance on the environmental risk assessment of genetically modified animals. *EFSA Journal*, 11(5), 3200.
<https://doi.org/10.2903/j.efsa.2013.3200>

Glover, K. A., Solberg, M. F., McGinnity, P., Hindar, K., Verspoor, E., Coulson, M. W., Hansen, M. M., Araki, H., Skaala, Ø., & Svåsand, T. (2017). Half a century of genetic interaction between farmed and wild Atlantic salmon: Status of knowledge and unanswered questions. *Fish and Fisheries*, 18(5), 890–927.
<https://doi.org/10.1111/faf.12214>

Gong, Z., Wan, H., Tay, T. L., Wang, H., Chen, M., & Yan, T. (2003). Development of transgenic fish for ornamental and bioreactor by strong expression of fluorescent proteins in the skeletal muscle. *Biochemical and Biophysical Research Communications*, 308(1), 58–63. [https://doi.org/10.1016/S0006-291X\(03\)01282-8](https://doi.org/10.1016/S0006-291X(03)01282-8)

Gutekunst, J., Andriantsoa, R., Falckenhayn, C., Hanna, K., Stein, W., Rasamy, J., & Lyko, F. (2018). Clonal genome evolution and rapid invasive spread of the marbled crayfish. *Nature Ecology & Evolution*, 2(3), 567–573. <https://doi.org/10.1038/s41559-018-0467-9>

Jha, P. (2010). Comparative study of aggressive behaviour in transgenic and wildtype zebrafish *Danio rerio* (Hamilton) and the flying barb *Esomus danricus* (Hamilton), and their susceptibility to predation by the snakehead *Channa striatus* (Bloch). *Italian Journal of Zoology*, 77(1), 102–109. <https://doi.org/10.1080/11250000802629463>

Leggatt, R. A., Hollo, T., Vandersteen, W. E., McFarlane, K., Goh, B., Prevost, J., & Devlin, R. H. (2014). Rearing in Seawater Mesocosms Improves the Spawning Performance of Growth Hormone Transgenic and Wild-Type Coho Salmon. *PLoS ONE*, 9(8), e105377. <https://doi.org/10.1371/journal.pone.0105377>

Lyko, F. (2017). The marbled crayfish (Decapoda: Cambaridae) represents an independent new species. *Zootaxa*, 4363(4), 544. <https://doi.org/10.11646/zootaxa.4363.4.6>

Norwegian Scientific Committee for Food and Environment (VKM), Hindar, K., Hole, L. R., Kausrud, K., Malmstrøm, M., Rimstad, E., Robertson, L., Sandlund, O. T., Thorstad, E. B., Vollset, K. W., de Boer, H., Eldegard, K., Järnegren, J., Kirkendall, L., Måren, I., Nielsen, A., Nilsen, E. B., Rueness, E., & Velle, G. (2020). Assessment of the risk to Norwegian biodiversity and aquaculture from pink salmon (*Oncorhynchus gorbuscha*) (VKM Report 2020:01).

<https://vkm.no/english/riskassessments/allpublications/assessmentoftheriskfromanincreaseofpinksalmoninnorway.4.303041af1695012160976b28.html>

Panskus, A. B., Miyazaki, J., & Then, C. (2020). RAGES subreport: Environmental risk assessment of genetically engineered crops that can persist and spontaneously propagate in the environment.

<https://www.testbiotech.org/en/content/rages-subreport-risk-assessment-next-generation-effects>

Secretariat of the International Plant Protection Convention. (2017). Pest risk analysis for quarantine pests (ISPM 11). <http://www.fao.org/3/a-j1302e.pdf>

Testbiotech. (2019). Testbiotech comment on the IUCN report “Genetic frontiers for conservation, an assessment of synthetic biology and biodiversity conservation.”

<https://www.testbiotech.org/content/testbiotech-comment-iucn-report-conservation-synthetic-biology>

Tuckett, Q. M., Ritch, J. L., Lawson, K. M., & Hill, J. E. (2017). Landscape-scale survey of non-native fishes near ornamental aquaculture facilities in Florida, USA. *Biological Invasions*, 19(1), 223–237.

<https://doi.org/10.1007/s10530-016-1275-2>

Online resources

BioTip pilot study: Genetic innovations as a trigger for phase transitions in the population dynamics of animals and plants (GeneTip): <https://www.genetip.de/en/biotip-pilot-study/>

Department of Fisheries and Aquatic Sciences at the University of Florida: <http://sfrc.ufl.edu/fish/>

DIYBiosphere: <https://sphere.diybio.org/>

Government of Canada - New substances risk assessment summaries: organisms:

<https://www.canada.ca/en/environment-climate-change/services/managing-pollution/evaluating-new-substances/biotechnology-living-organisms/risk-assessment-decisions.html>

Taikong Corporation: <http://www.azoo.com.tw/>

Testbiotech - comment on the IUCN report “Genetic frontiers for conservation, an assessment of synthetic biology and biodiversity conservation” <https://www.testbiotech.org/content/testbiotech-comment-iucn-report-conservation-synthetic-biology>

The Cuttlefish Project: <https://opensourcecuttlefish.com/>

The Odin - Frog Genetically Engineering kit: <https://www.the-odin.com/frog-ge-kit/>

IV. ADDITIONAL REFERENCES

Engineered gene drives

Adelman, Z. N., & Tu, Z. (2016). Control of Mosquito-Borne Infectious Diseases: Sex and Gene Drive. *Trends in Parasitology*, 32(3), 219–229. <https://doi.org/10.1016/j.pt.2015.12.003>

Akbari, O. S., Bellen, H. J., Bier, E., Bullock, S. L., Burt, A., Church, G. M., Cook, K. R., Duchek, P., Edwards, O. R., Esvelt, K. M., Gantz, V. M., Golic, K. G., Gratz, S. J., Harrison, M. M., Hayes, K. R., James, A. A., Kaufman, T. C., Knoblich, J., Malik, H. S., ... Wildonger, J. (2015). Safeguarding gene drive experiments in the laboratory. *Science*, 349(6251), 927–929. <https://doi.org/10.1126/science.aac7932>

Backus, G. A., & Delborne, J. A. (2019). Threshold-Dependent Gene Drives in the Wild: Spread, Controllability, and Ecological Uncertainty. *BioScience*. <https://doi.org/10.1093/biosci/biz098>

Baltzegar, J., Cavin Barnes, J., Elsensohn, J. E., Gutzmann, N., Jones, M. S., King, S., & Sudweeks, J. (2018). Anticipating complexity in the deployment of gene drive insects in agriculture. *Journal of Responsible Innovation*, 5(sup1), S81–S97. <https://doi.org/10.1080/23299460.2017.1407910>

Barrett, L. G., Legros, M., Kumaran, N., Glassop, D., Raghu, S., & Gardiner, D. M. (2019). Gene drives in plants: opportunities and challenges for weed control and engineered resilience. *Proceedings of the Royal Society B: Biological Sciences*, 286(1911), 20191515. <https://doi.org/10.1098/rspb.2019.1515>

Beaghton, A. K., Hammond, A., Nolan, T., Crisanti, A., & Burt, A. (2019). Gene drive for population genetic control: non-functional resistance and parental effects. *Proceedings of the Royal Society B: Biological Sciences*, 286(1914), 20191586. <https://doi.org/10.1098/rspb.2019.1586>

Buchman, A. B., Ivy, T., Marshall, J. M., Akbari, O. S., & Hay, B. A. (2018). Engineered Reciprocal Chromosome Translocations Drive High Threshold, Reversible Population Replacement in *Drosophila*. *ACS Synthetic Biology*, 7(5), 1359–1370. <https://doi.org/10.1021/acssynbio.7b00451>

Buchman, A., Marshall, J. M., Ostrovski, D., Yang, T., & Akbari, O. S. (2018). Synthetically engineered Medea gene drive system in the worldwide crop pest *Drosophila suzukii*. *Proceedings of the National Academy of Sciences*, 115(18), 4725–4730. <https://doi.org/10.1073/pnas.1713139115>

Bull, J. J., Remien, C. H., Gomulkiewicz, R., & Krone, S. M. (2019). Spatial structure undermines parasite suppression by gene drive cargo. *PeerJ*, 7, e7921. <https://doi.org/10.7717/peerj.7921>

Bull, J. J., Remien, C. H., & Krone, S. M. (2019). Gene-drive-mediated extinction is thwarted by population structure and evolution of sib mating. *Evolution, Medicine, and Public Health*, 2019(1), 66–81. <https://doi.org/10.1093/emph/eoz014>

Burt, A., & Crisanti, A. (2018). Gene Drive: Evolved and Synthetic. *ACS Chemical Biology*, 13(2), 343–346. <https://doi.org/10.1021/acschembio.7b01031>

- Champer, J., Buchman, A., & Akbari, O. S. (2016). Cheating evolution: engineering gene drives to manipulate the fate of wild populations. *Nature Reviews Genetics*, 17(3), 146–159.
<https://doi.org/10.1038/nrg.2015.34>
- Champer, J., Chung, J., Lee, Y. L., Liu, C., Yang, E., Wen, Z., Clark, A. G., & Messer, P. W. (2019). Molecular safeguarding of CRISPR gene drive experiments. *eLife*, 8. <https://doi.org/10.7554/eLife.41439>
- Champer, J., Liu, J., Oh, S. Y., Reeves, R., Luthra, A., Oakes, N., Clark, A. G., & Messer, P. W. (2018). Reducing resistance allele formation in CRISPR gene drive. *Proceedings of the National Academy of Sciences*, 115(21), 5522–5527. <https://doi.org/10.1073/pnas.1720354115>
- Champer, J., Reeves, R., Oh, S. Y., Liu, C., Liu, J., Clark, A. G., & Messer, P. W. (2017). Novel CRISPR/Cas9 gene drive constructs reveal insights into mechanisms of resistance allele formation and drive efficiency in genetically diverse populations. *PLOS Genetics*, 13(7), e1006796.
<https://doi.org/10.1371/journal.pgen.1006796>
- Champer, J., Wen, Z., Luthra, A., Reeves, R., Chung, J., Liu, C., Lee, Y. L., Liu, J., Yang, E., Messer, P. W., & Clark, A. G. (2019). CRISPR Gene Drive Efficiency and Resistance Rate Is Highly Heritable with No Common Genetic Loci of Large Effect. *Genetics*, genetics.302037.2019.
<https://doi.org/10.1534/genetics.119.302037>
- Dhole, S., Lloyd, A. L., & Gould, F. (2019). Tethered homing gene drives: A new design for spatially restricted population replacement and suppression. *Evolutionary Applications*, 12(8), 1688–1702.
<https://doi.org/10.1111/eva.12827>
- Dhole, S., Vella, M. R., Lloyd, A. L., & Gould, F. (2018). Invasion and migration of spatially self-limiting gene drives: A comparative analysis. *Evolutionary Applications*, 11(5), 794–808.
<https://doi.org/10.1111/eva.12583>
- DiCarlo, J. E., Chavez, A., Dietz, S. L., Esvelt, K. M., & Church, G. M. (2015). Safeguarding CRISPR-Cas9 gene drives in yeast. *Nature Biotechnology*, 33(12), 1250–1255. <https://doi.org/10.1038/nbt.3412>
- Drury, D. W., Dapper, A. L., Siniard, D. J., Zentner, G. E., & Wade, M. J. (2017). CRISPR/Cas9 gene drives in genetically variable and nonrandomly mating wild populations. *Science Advances*, 3(5), e1601910. <https://doi.org/10.1126/sciadv.1601910>
- Eckhoff, P. A., Wenger, E. A., Godfray, H. C. J., & Burt, A. (2017). Impact of mosquito gene drive on malaria elimination in a computational model with explicit spatial and temporal dynamics. *Proceedings of the National Academy of Sciences*, 114(2), E255–E264. <https://doi.org/10.1073/pnas.1611064114>
- Edgington, M. P., & Alphey, L. S. (2019). Modeling the mutation and reversal of engineered underdominance gene drives. *Journal of Theoretical Biology*, 479, 14–21.
<https://doi.org/10.1016/j.jtbi.2019.06.024>
- Esvelt, K. M., & Gemmell, N. J. (2017). Conservation demands safe gene drive. *PLOS Biology*, 15(11), e2003850. <https://doi.org/10.1371/journal.pbio.2003850>
- Frieß, J. L., von Gleich, A., & Giese, B. (2019). Gene drives as a new quality in GMO releases—a comparative technology characterization. *PeerJ*, 7, e6793. <https://doi.org/10.7717/peerj.6793>

Godwin, J., Serr, M., Barnhill-Dilling, S. K., Blondel, D. V., Brown, P. R., Campbell, K., Delborne, J., Lloyd, A. L., Oh, K. P., Prowse, T. A. A., Saah, R., & Thomas, P. (2019). Rodent gene drives for conservation: opportunities and data needs. *Proceedings of the Royal Society B: Biological Sciences*, 286(1914), 20191606. <https://doi.org/10.1098/rspb.2019.1606>

Grunwald, H. A., Gantz, V. M., Poplawski, G., Xu, X.-R. S., Bier, E., & Cooper, K. L. (2019). Super-Mendelian inheritance mediated by CRISPR–Cas9 in the female mouse germline. *Nature*, 566(7742), 105–109. <https://doi.org/10.1038/s41586-019-0875-2>

Hammond, A. M., Kyrou, K., Bruttini, M., North, A., Galizi, R., Karlsson, X., Kranjc, N., Carpi, F. M., D'Aurizio, R., Crisanti, A., & Nolan, T. (2017). The creation and selection of mutations resistant to a gene drive over multiple generations in the malaria mosquito. *PLOS Genetics*, 13(10), e1007039. <https://doi.org/10.1371/journal.pgen.1007039>

Hammond, A., Galizi, R., Kyrou, K., Simoni, A., Siniscalchi, C., Katsanos, D., Gribble, M., Baker, D., Marois, E., Russell, S., Burt, A., Windbichler, N., Crisanti, A., & Nolan, T. (2016). A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*. *Nature Biotechnology*, 34(1), 78–83. <https://doi.org/10.1038/nbt.3439>

Hayes, K. R., Hosack, G. R., Dana, G. V., Foster, S. D., Ford, J. H., Thresher, R., Ickowicz, A., Peel, D., Tizard, M., De Barro, P., Strive, T., & Dambacher, J. M. (2018). Identifying and detecting potentially adverse ecological outcomes associated with the release of gene-drive modified organisms. *Journal of Responsible Innovation*, 5(sup1), S139–S158. <https://doi.org/10.1080/23299460.2017.1415585>

Heffel, M. G., & Finnigan, G. C. (2019). Mathematical modeling of self-contained CRISPR gene drive reversal systems. *Scientific Reports*, 9(1), 20050. <https://doi.org/10.1038/s41598-019-54805-8>

Holman, L. (2019). Evolutionary simulations of Z-linked suppression gene drives. *Proceedings of the Royal Society B: Biological Sciences*, 286(1912), 20191070. <https://doi.org/10.1098/rspb.2019.1070>

James, S., Collins, F. H., Welkhoff, P. A., Emerson, C., Godfray, H. C., Gottlieb, M., Greenwood, B., Lindsay, S. W., Mbogo, C. M., Okumu, F. O., Quemada, H., Savadogo, M., Singh, J. A., Tountas, K. H., & Toure, Y. T. (2018). Pathway to deployment of gene drive mosquitoes as a potential biocontrol tool for elimination of malaria in sub-Saharan Africa: Recommendations of a scientific working group. *American Journal of Tropical Medicine and Hygiene*, 98(Suppl 6), 1–49. <https://doi.org/10.4269/ajtmh.18-0083>

Kandul, N. P., Liu, J., Sanchez C., H. M., Wu, S. L., Marshall, J. M., & Akbari, O. S. (2019). Transforming insect population control with precision guided sterile males with demonstration in flies. *Nature Communications*, 10(1), 84. <https://doi.org/10.1038/s41467-018-07964-7>

KaramiNejadRanjbar, M., Eckermann, K. N., Ahmed, H. M. M., Sánchez C., H. M., Dippel, S., Marshall, J. M., & Wimmer, E. A. (2018). Consequences of resistance evolution in a Cas9-based sex conversion-suppression gene drive for insect pest management. *Proceedings of the National Academy of Sciences*, 115(24), 6189–6194. <https://doi.org/10.1073/pnas.1713825115>

Kuzma, J. (2019). Procedurally Robust Risk Assessment Framework for Novel Genetically Engineered Organisms and Gene Drives. *Regulation & Governance*, rego.12245. <https://doi.org/10.1111/rego.12245>

Kyrou, K., Hammond, A. M., Galizi, R., Kranjc, N., Burt, A., Beaghton, A. K., Nolan, T., & Crisanti, A. (2018). A CRISPR–Cas9 gene drive targeting doublesex causes complete population suppression in caged

Anopheles gambiae mosquitoes. *Nature Biotechnology*, 36(11), 1062–1066.
<https://doi.org/10.1038/nbt.4245>

Leftwich, P. T., Edgington, M. P., Harvey-Samuel, T., Carabajal Paladino, L. Z., Norman, V. C., & Alphey, L. (2018). Recent advances in threshold-dependent gene drives for mosquitoes. *Biochemical Society Transactions*, 46(5), 1203–1212. <https://doi.org/10.1042/BST20180076>

Leitschuh, C. M., Kanavy, D., Backus, G. A., Valdez, R. X., Serr, M., Pitts, E. A., Threadgill, D., & Godwin, J. (2018). Developing gene drive technologies to eradicate invasive rodents from islands. *Journal of Responsible Innovation*, 5(sup1), S121–S138. <https://doi.org/10.1080/23299460.2017.1365232>

Li, M., Yang, T., Kandul, N. P., Bui, M., Gamez, S., Raban, R., Bennett, J., Sánchez C, H. M., Lanzaro, G. C., Schmidt, H., Lee, Y., Marshall, J. M., & Akbari, O. S. (2020). Development of a confinable gene drive system in the human disease vector *Aedes aegypti*. *eLife*, 9. <https://doi.org/10.7554/eLife.51701>

Macias, V., Ohm, J., & Rasgon, J. (2017). Gene Drive for Mosquito Control: Where Did It Come from and Where Are We Headed? *International Journal of Environmental Research and Public Health*, 14(9), 1006. <https://doi.org/10.3390/ijerph14091006>

Maier, T., Wheeler, N. J., Namigai, E. K. O., Tycko, J., Grawelle, R. E., Woldeamanuel, Y., Klohe, K., Perez-Saez, J., Sokolow, S. H., De Leo, G. A., Yoshino, T. P., Zamanian, M., & Reinhard-Rupp, J. (2019). Gene drives for schistosomiasis transmission control. *PLOS Neglected Tropical Diseases*, 13(12), e0007833. <https://doi.org/10.1371/journal.pntd.0007833>

Manser, A., Cornell, S. J., Sutter, A., Blondel, D. V., Serr, M., Godwin, J., & Price, T. A. R. (2019). Controlling invasive rodents via synthetic gene drive and the role of polyandry. *Proceedings of the Royal Society B: Biological Sciences*, 286(1909), 20190852. <https://doi.org/10.1098/rspb.2019.0852>

Marshall, J. M., & Akbari, O. S. (2018). Can CRISPR-Based Gene Drive Be Confined in the Wild? A Question for Molecular and Population Biology. *ACS Chemical Biology*, 13(2), 424–430. <https://doi.org/10.1021/acschembio.7b00923>

McFarlane, G. R., Whitelaw, C. B. A., & Lillico, S. G. (2018). CRISPR-Based Gene Drives for Pest Control. *Trends in Biotechnology*, 36(2), 130–133. <https://doi.org/10.1016/j.tibtech.2017.10.001>

Mitchell, P. D., Brown, Z., & McRoberts, N. (2018). Economic issues to consider for gene drives. *Journal of Responsible Innovation*, 5(sup1), S180–S202. <https://doi.org/10.1080/23299460.2017.1407914>

Moro, D., Byrne, M., Kennedy, M., Campbell, S., & Tizard, M. (2018). Identifying knowledge gaps for gene drive research to control invasive animal species: The next CRISPR step. *Global Ecology and Conservation*, 13, e00363. <https://doi.org/10.1016/j.gecco.2017.e00363>

Nash, A., Urdaneta, G. M., Beaghton, A. K., Hoermann, A., Papathanos, P. A., Christophides, G. K., & Windbichler, N. (2019). Integral gene drives for population replacement. *Biology Open*, 8(1), bio037762. <https://doi.org/10.1242/bio.037762>

Neve, P. (2018). Gene drive systems: do they have a place in agricultural weed management? *Pest Management Science*, 74(12), 2671–2679. <https://doi.org/10.1002/ps.5137>

Noble, C., Adlam, B., Church, G. M., Esvelt, K. M., & Nowak, M. A. (2018). Current CRISPR gene drive systems are likely to be highly invasive in wild populations. *eLife*, 7. <https://doi.org/10.7554/eLife.33423>

Noble, C., Min, J., Olejarz, J., Buchthal, J., Chavez, A., Smidler, A. L., DeBenedictis, E. A., Church, G. M., Nowak, M. A., & Esvelt, K. M. (2019). Daisy-chain gene drives for the alteration of local populations. *Proceedings of the National Academy of Sciences*, 201716358. <https://doi.org/10.1073/pnas.1716358116>

Oberhofer, G., Ivy, T., & Hay, B. A. (2018). Behavior of homing endonuclease gene drives targeting genes required for viability or female fertility with multiplexed guide RNAs. *Proceedings of the National Academy of Sciences*, 115(40), E9343–E9352. <https://doi.org/10.1073/pnas.1805278115>

Oberhofer, G., Ivy, T., & Hay, B. A. (2019). Cleave and Rescue, a novel selfish genetic element and general strategy for gene drive. *Proceedings of the National Academy of Sciences*, 116(13), 6250–6259. <https://doi.org/10.1073/pnas.1816928116>

Price, T. A. R., Verspoor, R., & Wedell, N. (2019). Ancient gene drives: an evolutionary paradox. *Proceedings of the Royal Society B: Biological Sciences*, 286(1917), 20192267. <https://doi.org/10.1098/rspb.2019.2267>

Prowse, T. A. A., Cassey, P., Ross, J. V., Pfitzner, C., Wittmann, T. A., & Thomas, P. (2017). Dodging silver bullets: good CRISPR gene-drive design is critical for eradicating exotic vertebrates. *Proceedings of the Royal Society B: Biological Sciences*, 284(1860), 20170799. <https://doi.org/10.1098/rspb.2017.0799>

Prowse, T. A., Adikusuma, F., Cassey, P., Thomas, P., & Ross, J. V. (2019). A Y-chromosome shredding gene drive for controlling pest vertebrate populations. *eLife*, 8. <https://doi.org/10.7554/eLife.41873>

Rode, N. O., Estoup, A., Bourguet, D., Courtier-Orgogozo, V., & Débarre, F. (2019). Population management using gene drive: molecular design, models of spread dynamics and assessment of ecological risks. *Conservation Genetics*, 20(4), 671–690. <https://doi.org/10.1007/s10592-019-01165-5>

Simon, S., Otto, M., & Engelhard, M. (2018). Synthetic gene drive: between continuity and novelty. *EMBO Reports*, 19(5), e45760. <https://doi.org/10.15252/embr.201845760>

Snow, A. A. (2019). Genetically Engineering Wild Mice to Combat Lyme Disease: An Ecological Perspective. *BioScience*, 69(9), 746–756. <https://doi.org/10.1093/biosci/biz080>

Unckless, R. L., Clark, A. G., & Messer, P. W. (2017). Evolution of Resistance Against CRISPR/Cas9 Gene Drive. *Genetics*, 205(2), 827–841. <https://doi.org/10.1534/genetics.116.197285>

Wedell, N., Price, T. A. R., & Lindholm, A. K. (2019). Gene drive: progress and prospects. *Proceedings of the Royal Society B: Biological Sciences*, 286(1917), 20192709. <https://doi.org/10.1098/rspb.2019.2709>

Wu, B., Luo, L., & Gao, X. J. (2016). Cas9-triggered chain ablation of cas9 as a gene drive brake. *Nature Biotechnology*, 34(2), 137–138. <https://doi.org/10.1038/nbt.3444>

Yan, Y., & Finnigan, G. C. (2018). Development of a multi-locus CRISPR gene drive system in budding yeast. *Scientific Reports*, 8(1), 17277. <https://doi.org/10.1038/s41598-018-34909-3>

Living modified fish

Anderson, J. L., Mulligan, T. S., Shen, M.-C., Wang, H., Scahill, C. M., Tan, F. J., Du, S. J., Busch-Nentwich, E. M., & Farber, S. A. (2017). mRNA processing in mutant zebrafish lines generated by chemical and CRISPR-mediated mutagenesis produces unexpected transcripts that escape nonsense-mediated decay. *PLOS Genetics*, 13(11), e1007105. <https://doi.org/10.1371/journal.pgen.1007105>

Crossin, G. T., & Devlin, R. H. (2017). Predation, metabolic priming and early life-history rearing environment affect the swimming capabilities of growth hormone transgenic rainbow trout. *Biology Letters*, 13(8), 20170279. <https://doi.org/10.1098/rsbl.2017.0279>

Devlin, R. H., Sundström, L. F., & Leggatt, R. A. (2015). Assessing Ecological and Evolutionary Consequences of Growth-Accelerated Genetically Engineered Fishes. *BioScience*, 65(7), 685–700. <https://doi.org/10.1093/biosci/biv068>

Hill, J. E., Lawson, L. L., & Hardin, S. (2014). Assessment of the Risks of Transgenic Fluorescent Ornamental Fishes to the United States Using the Fish Invasiveness Screening Kit (FISK). *Transactions of the American Fisheries Society*, 143(3), 817–829. <https://doi.org/10.1080/00028487.2014.880741>

Ishikawa, T., Ansai, S., Kinoshita, M., & Mori, K. (2018). A Collection of Transgenic Medaka Strains for Efficient Site-Directed Transgenesis Mediated by phiC31 Integrase. *G3: Genes/Genomes/Genetics*, 8(8), 2585–2593. <https://doi.org/10.1534/g3.118.200130>

Jiang, X.-Y., Huang, C.-X., Zhong, S.-S., Sun, C.-F., & Zou, S.-M. (2017). Transgenic overexpression of follistatin 2 in blunt snout bream results in increased muscle mass caused by hypertrophy. *Aquaculture*, 468, 442–450. <https://doi.org/10.1016/j.aquaculture.2016.11.014>

Kim, J.-H., Leggatt, R. A., Chan, M., Volkoff, H., & Devlin, R. H. (2015). Effects of chronic growth hormone overexpression on appetite-regulating brain gene expression in coho salmon. *Molecular and Cellular Endocrinology*, 413, 178–188. <https://doi.org/10.1016/j.mce.2015.06.024>

Kim, J.-H., Macqueen, D. J., Winton, J. R., Hansen, J. D., Park, H., & Devlin, R. H. (2019). Effect of growth rate on transcriptomic responses to immune stimulation in wild-type, domesticated, and GH-transgenic coho salmon. *BMC Genomics*, 20(1), 1024. <https://doi.org/10.1186/s12864-019-6408-4>

Leggatt, R. A., Biagi, C. A., Sakhrani, D., Dominelli, R., Eliason, E. J., Farrell, A. P., & Devlin, R. H. (2017). Fitness component assessments of wild-type and growth hormone transgenic coho salmon reared in seawater mesocosms. *Aquaculture*, 473, 31–42. <https://doi.org/10.1016/j.aquaculture.2017.01.022>

Leggatt, R. A., & Devlin, R. H. (2019). Fluorescent protein transgenesis has varied effects on behaviour and cold tolerance in a tropical fish (*Gymnocranius ternetzi*): implications for risk assessment. *Fish Physiology and Biochemistry*. <https://doi.org/10.1007/s10695-019-00725-3>

Leggatt, R. A., Sundström, L. F., Woodward, K., & Devlin, R. H. (2017). Growth-Enhanced Transgenic Coho Salmon (*Oncorhynchus kisutch*) Strains Have Varied Success in Simulated Streams: Implications for Risk Assessment. *PLOS ONE*, 12(1), e0169991. <https://doi.org/10.1371/journal.pone.0169991>

Li, H., Su, B., Qin, G., Ye, Z., Alsaifi, A., Perera, D., Shang, M., Odin, R., Vo, K., Drescher, D., Robinson, D., Zhang, D., Abass, N., & Dunham, R. (2017). Salt Sensitive Tet-Off-Like Systems to Knockdown Primordial Germ Cell Genes for Repressible Transgenic Sterilization in Channel Catfish, *Ictalurus punctatus*. *Marine Drugs*, 15(6), 155. <https://doi.org/10.3390/md15060155>

Li, H., Su, B., Qin, G., Ye, Z., Elaswad, A., Alsaqufi, A., Perera, D. A., Qin, Z., Odin, R., Vo, K., Drescher, D., Robinson, D., Dong, S., Zhang, D., Shang, M., Abass, N., Das, S. K., Bangs, M., & Dunham, R. A. (2018). Repressible Transgenic Sterilization in Channel Catfish, *Ictalurus punctatus*, by Knockdown of Primordial Germ Cell Genes with Copper-Sensitive Constructs. *Marine Biotechnology*, 20(3), 324–342. <https://doi.org/10.1007/s10126-018-9819-3>

Luo, L., Huang, R., Zhang, A., Yang, C., Chen, L., Zhu, D., Li, Y., He, L., Liao, L., Zhu, Z., & Wang, Y. (2018). Selection of growth-related genes and dominant genotypes in transgenic Yellow River carp *Cyprinus carpio* L. *Functional & Integrative Genomics*, 18(4), 425–437. <https://doi.org/10.1007/s10142-018-0597-9>

Mei, J., & Gui, J.-F. (2015). Genetic basis and biotechnological manipulation of sexual dimorphism and sex determination in fish. *Science China Life Sciences*, 58(2), 124–136. <https://doi.org/10.1007/s11427-014-4797-9>

Moreau, D. T. R., Gamperl, A. K., Fletcher, G. L., & Fleming, I. A. (2014). Delayed Phenotypic Expression of Growth Hormone Transgenesis during Early Ontogeny in Atlantic Salmon (*Salmo salar*)? *PLoS ONE*, 9(4), e95853. <https://doi.org/10.1371/journal.pone.0095853>

Noble, S., Saxena, V., Ekker, M., & Devlin, R. (2017). Expression of Thiaminase in Zebrafish (*Danio rerio*) is Lethal and Has Implications for Use as a Biocontainment Strategy in Aquaculture and Invasive Species. *Marine Biotechnology*, 19(6), 563–569. <https://doi.org/10.1007/s10126-017-9776-2>

Oke, K. B., Westley, P. A. H., Moreau, D. T. R., & Fleming, I. A. (2013). Hybridization between genetically modified Atlantic salmon and wild brown trout reveals novel ecological interactions. *Proceedings of the Royal Society B: Biological Sciences*, 280(1763), 20131047. <https://doi.org/10.1098/rspb.2013.1047>

Pauliny, A., Devlin, R. H., Johnsson, J. I., & Blomqvist, D. (2015). Rapid growth accelerates telomere attrition in a transgenic fish. *BMC Evolutionary Biology*, 15(1), 159. <https://doi.org/10.1186/s12862-015-0436-8>

Pawar, N., Gireesh-Babu, P., Sabnis, S., Rasal, K., Murthy, R., Zaidi, S. G. S., Sivasubbu, S., & Chaudhari, A. (2016). Development of a fluorescent transgenic zebrafish biosensor for sensing aquatic heavy metal pollution. *Transgenic Research*, 25(5), 617–627. <https://doi.org/10.1007/s11248-016-9959-z>

Qin, Z., Li, Y., Su, B., Cheng, Q., Ye, Z., Perera, D. A., Fobes, M., Shang, M., & Dunham, R. A. (2016). Editing of the Luteinizing Hormone Gene to Sterilize Channel Catfish, *Ictalurus punctatus*, Using a Modified Zinc Finger Nuclease Technology with Electroporation. *Marine Biotechnology*, 18(2), 255–263. <https://doi.org/10.1007/s10126-016-9687-7>

Rossi, A., Kontarakis, Z., Gerri, C., Nolte, H., Hölder, S., Krüger, M., & Stainier, D. Y. R. (2015). Genetic compensation induced by deleterious mutations but not gene knockdowns. *Nature*, 524(7564), 230–233. <https://doi.org/10.1038/nature14580>

Shiraki, T., & Kawakami, K. (2018). A tRNA-based multiplex sgRNA expression system in zebrafish and its application to generation of transgenic albino fish. *Scientific Reports*, 8(1), 13366. <https://doi.org/10.1038/s41598-018-31476-5>

Straume, A. H., Kjærner-Semb, E., Ove Skaftnesmo, K., Güralp, H., Kleppe, L., Wargelius, A., & Edvardsen, R. B. (2020). Indel locations are determined by template polarity in highly efficient in vivo

CRISPR/Cas9-mediated HDR in Atlantic salmon. *Scientific Reports*, 10(1), 409.

<https://doi.org/10.1038/s41598-019-57295-w>

Su, B.-C., Lai, Y.-W., Chen, J.-Y., & Pan, C.-Y. (2018). Transgenic expression of tilapia piscidin 3 (TP3) in zebrafish confers resistance to *Streptococcus agalactiae*. *Fish & Shellfish Immunology*, 74, 235–241.

<https://doi.org/10.1016/j.fsi.2018.01.001>

Sundström, L. F., Löhmus, M., & Devlin, R. H. (2016). Gene-environment interactions influence feeding and anti-predator behavior in wild and transgenic coho salmon. *Ecological Applications*, 26(1), 67–76.

<https://doi.org/10.1890/15-0252>

Sundström, L. F., & Devlin, R. H. (2011). Increased intrinsic growth rate is advantageous even under ecologically stressful conditions in coho salmon (*Oncorhynchus kisutch*). *Evolutionary Ecology*, 25(2), 447–460. <https://doi.org/10.1007/s10682-010-9406-1>

Sundström, L. F., & Devlin, R. H. (2015). Ecological implications of genetically modified fishes in freshwater fisheries, with a focus on salmonids. In *Freshwater Fisheries Ecology* (pp. 594–615). John Wiley & Sons, Ltd. <https://doi.org/10.1002/9781118394380.ch46>

Tonelli, F. M. P., Lacerda, S. M. S. N., Tonelli, F. C. P., Costa, G. M. J., de França, L. R., & Resende, R. R. (2017). Progress and biotechnological prospects in fish transgenesis. *Biotechnology Advances*, 35(6), 832–844. <https://doi.org/10.1016/j.biotechadv.2017.06.002>

Trump, B. D., Foran, C., Rycroft, T., Wood, M. D., Bandolin, N., Cains, M., Cary, T., Crocker, F., Friedenberg, N. A., Gurian, P., Hamilton, K., Hoover, J. J., Meyer, C., Pokrzywinski, K., Ritterson, R., Schulte, P., Warner, C., Perkins, E., & Linkov, I. (2018). Development of community of practice to support quantitative risk assessment for synthetic biology products: contaminant bioremediation and invasive carp control as cases. *Environment Systems and Decisions*, 38(4), 517–527.

<https://doi.org/10.1007/s10669-018-9710-9>

Vandersteen, W. E., Leggatt, R., Sundström, L. F., & Devlin, R. H. (2019). Importance of Experimental Environmental Conditions in Estimating Risks and Associated Uncertainty of Transgenic Fish Prior to Entry into Nature. *Scientific Reports*, 9(1), 406. <https://doi.org/10.1038/s41598-018-35826-1>

Wei, Y., Huang, L., Cao, J., Wang, C., & Yan, J. (2018). Dietary Safety Assessment of Flk1-Transgenic Fish. *Frontiers in Physiology*, 9. <https://doi.org/10.3389/fphys.2018.00008>

White, S. L., Volkoff, H., & Devlin, R. H. (2016). Regulation of feeding behavior and food intake by appetite-regulating peptides in wild-type and growth hormone-transgenic coho salmon. *Hormones and Behavior*, 84, 18–28. <https://doi.org/10.1016/j.ybeh.2016.04.005>

Wu, S.-H., Lin, H.-J., Lin, W.-F., Wu, J.-L., & Gong, H.-Y. (2018). A potent tilapia secreted granulin peptide enhances the survival of transgenic zebrafish infected by *Vibrio vulnificus* via modulation of innate immunity. *Fish & Shellfish Immunology*, 75, 74–90. <https://doi.org/10.1016/j.fsi.2018.01.044>

Yeh, Y.-C., Kinoshita, M., Ng, T. H., Chang, Y.-H., Maekawa, S., Chiang, Y.-A., Aoki, T., & Wang, H.-C. (2017). Using CRISPR/Cas9-mediated gene editing to further explore growth and trade-off effects in myostatin-mutated F4 medaka (*Oryzias latipes*). *Scientific Reports*, 7(1), 11435.

<https://doi.org/10.1038/s41598-017-09966-9>

Yin, L., Maddison, L. A., Li, M., Kara, N., LaFave, M. C., Varshney, G. K., Burgess, S. M., Patton, J. G., & Chen, W. (2015). Multiplex Conditional Mutagenesis Using Transgenic Expression of Cas9 and sgRNAs. *Genetics*, 200(2), 431–441. <https://doi.org/10.1534/genetics.115.176917>

Zhang, L., Gozlan, R. E., Li, Z., Liu, J., Zhang, T., Hu, W., & Zhu, Z. (2014). Rapid growth increases intrinsic predation risk in genetically modified *Cyprinus carpio*: implications for environmental risk. *Journal of Fish Biology*, 84(5), 1527–1538. <https://doi.org/10.1111/jfb.12381>

Zhang, X., Pang, S., Liu, C., Wang, H., Ye, D., Zhu, Z., & Sun, Y. (2019). A Novel Dietary Source of EPA and DHA: Metabolic Engineering of an Important Freshwater Species—Common Carp by fat1-Transgenesis. *Marine Biotechnology*, 21(2), 171–185. <https://doi.org/10.1007/s10126-018-9868-7>

Zhang, Y., Chen, J., Cui, X., Luo, D., Xia, H., Dai, J., Zhu, Z., & Hu, W. (2015). A controllable on-off strategy for the reproductive containment of fish. *Scientific Reports*, 5(1), 7614. <https://doi.org/10.1038/srep07614>

Zhu, T. B., Zhang, L. H., Zhang, T. L., Wang, Y. P., Hu, W., Ringø, E., & Zhu, Z. Y. (2017). Effects of sustained predation by fast-growing transgenic common carp (*Cyprinus carpio* Linnaeus, 1758) on gastropods in artificial environments. *Journal of Applied Ichthyology*, 33(1), 22–28. <https://doi.org/10.1111/jai.13130>

Zhu, T., Zhang, L., Zhang, T., Wang, Y., Hu, W., Olsen, R. E., & Zhu, Z. (2018). Preliminarily study on the maximum handling size, prey size and species selectivity of growth hormone transgenic and non-transgenic common carp *Cyprinus carpio* when foraging on gastropods. *Journal of Oceanology and Limnology*, 36(4), 1425–1433. <https://doi.org/10.1007/s00343-018-7045-5>

Policies, reports, perspectives and opinions

Gene drive overdrive. (2015). *Nature Biotechnology*, 33(10), 1019–1021. <https://doi.org/10.1038/nbt.3361>

African Centre for Biodiversity. (2018). *Critique of African Union and NEPAD's positions on gene drive mosquitoes for Malaria elimination* (p. 18).

https://www.acbio.org.za/sites/default/files/documents/Critique_of_African_Union_and_NEPADs_positions_on_gene_drive_mosquitoes_for_Malaria_elimination.pdf

African Union, & NEPAD. (2018). *Gene drives for malaria control and elimination in Africa*. www.nepad.org

Alphey, L. (2016). Can CRISPR-Cas9 gene drives curb malaria? *Nature Biotechnology*, 34(2), 149–150. <https://doi.org/10.1038/nbt.3473>

Australian Academy of Science. (2017). *Synthetic Gene Drives in Australia: Implications of Emerging Technologies*. <https://www.science.org.au/support/analysis/reports/synthetic-gene-drives-australia-implications-emerging-technologies>

Bezerra, L. A. V., Freitas, M. O., Daga, V. S., Occhi, T. V. T., Faria, L., Costa, A. P. L., Padial, A. A., Prodocimo, V., & Vitule, J. R. S. (2019). A network meta-analysis of threats to South American fish biodiversity. *Fish and Fisheries*, faf.12365. <https://doi.org/10.1111/faf.12365>

Brossard, D., Belluck, P., Gould, F., & Wirz, C. D. (2019). Promises and perils of gene drives: Navigating the communication of complex, post-normal science. *Proceedings of the National Academy of Sciences*, 116(16), 7692–7697. <https://doi.org/10.1073/pnas.1805874115>

Bruce, A. B. (2017). Frankenfish or Fish to Feed the World? Scientism and Biotechnology Regulatory Policy. *Rural Sociology*, 82(4), 628–663. <https://doi.org/10.1111/ruso.12146>

Cotter, J., Kawall, K., & Then, C. (2020). *New genetic engineering technologies. Report of the results from the RAGES project 2016-2019.* http://www.testbiotech.org/projekt_rages

Courtier- Orgogozo, V., Morizot, B., & Boëte, C. (2017). Agricultural pest control with CRISPR- based gene drive: time for public debate. *EMBO Reports*, 18(6), 878–880. <https://doi.org/10.15252/embr.201744205>

Critical Scientists Switzerland, European Network of Scientists for Social and Environmental Responsibility, & Vereinigung Deutscher Wissenschaftler. (2019). *Gene drives: A report on their science, applications, social aspects, ethics and regulations.* <https://genedrives.ch/report>

Devos, Y., Craig, W., Devlin, R. H., Ippolito, A., Leggatt, R. A., Romeis, J., Shaw, R., Svendsen, C., & Topping, C. J. (2019). Using problem formulation for fit- for- purpose pre- market environmental risk assessments of regulated stressors. *EFSA Journal*, 17. <https://doi.org/10.2903/j.efsa.2019.e170708>

Esveld, K. M., Smidler, A. L., Catteruccia, F., & Church, G. M. (2014). Concerning RNA-guided gene drives for the alteration of wild populations. *eLife*, 3. <https://doi.org/10.7554/eLife.03401>

ETC Group. (2018). *Forcing the Farm: How gene drive organisms could entrench industrial agriculture and threaten food sovereignty.* www.etcgroup.com

George, D. R., Kuiken, T., & Delborne, J. A. (2019). Articulating ‘free, prior and informed consent’ (FPIC) for engineered gene drives. *Proceedings of the Royal Society B: Biological Sciences*, 286(1917), 20191484. <https://doi.org/10.1098/rspb.2019.1484>

Godwin, J., Serr, M., Barnhill-Dilling, S. K., Blondel, D. V., Brown, P. R., Campbell, K., Delborne, J., Lloyd, A. L., Oh, K. P., Prowse, T. A. A., Saah, R., & Thomas, P. (2019). Rodent gene drives for conservation: opportunities and data needs. *Proceedings of the Royal Society B: Biological Sciences*, 286(1914), 20191606. <https://doi.org/10.1098/rspb.2019.1606>

Haut Conseil des Biotechnologies. (2017). *Scientific Opinion in response to the referral of 12 October 2015 concerning the use of genetically modified mosquitoes for vector control* (p. 142). <http://www.hautconseildesbiotechnologies.fr/en/avis/avis-relatif-a-lutilisation-moustiques-gm-dans-cadre-lutte-antivectorielle>

Hokanson, K. E. (2019). When Policy Meets Practice: The Dilemma for Guidance on Risk Assessment Under the Cartagena Protocol on Biosafety. *Frontiers in Bioengineering and Biotechnology*, 7. <https://doi.org/10.3389/fbioe.2019.00082>

James, S., & Tountas, K. (2018). Using Gene Drive Technologies to Control Vector-Borne Infectious Diseases. *Sustainability*, 10(12), 4789. <https://doi.org/10.3390/su10124789>

Joks, S., & Law, J. (2017). Sámi salmon, state salmon: TEK, technoscience and care. *The Sociological Review*, 65(2_suppl), 150–171. <https://doi.org/10.1177/0081176917710428>

Meghani, Z. (2019). Autonomy of Nations and Indigenous Peoples and the Environmental Release of Genetically Engineered Animals with Gene Drives. *Global Policy*, 10(4), 554–568. <https://doi.org/10.1111/1758-5899.12699>

Morris, M. C. (2019). Predator Free New Zealand and the ‘War’ on Pests: Is it a just War? *Journal of Agricultural and Environmental Ethics*. <https://doi.org/10.1007/s10806-019-09815-x>

Myskja, B. K., & Myhr, A. I. (2019). 36. Moral limits to genome editing of farmed salmon. *Sustainable Governance and Management of Food Systems*, 261–266. https://doi.org/10.3920/978-90-8686-892-6_36

National Academies of Sciences Engineering and Medicine. (2016). *Gene Drives on the Horizon*. National Academies Press. <https://doi.org/10.17226/23405>

Norwegian Scientific Committee for Food and Environment (VKM), Hindar, K., Hole, L. R., Kausrud, K., Malmstrøm, M., Rimstad, E., Robertson, L., Sandlund, O. T., Thorstad, E. B., Vollset, K. W., de Boer, H., Eldegard, K., Järnegren, J., Kirkendall, L., Måren, I., Nielsen, A., Nilsen, E. B., Rueness, E., & Velle, G. (2020). *Assessment of the risk to Norwegian biodiversity and aquaculture from pink salmon (Oncorhynchus gorbuscha) (VKM Report 2020:01)*. <https://vkm.no/english/riskassessments/allpublications/assessmentoftheriskfromanincreaseofpinksalmoninnorway.4.303041af1695012160976b28.html>

OECD. (2019). *Safety of novel foods and feeds and on the harmonisation of regulatory oversight in biotechnology*. <http://www.oecd.org/chemicalsafety/biotrack/oecdandrisksafetyassessmentinmodernbiotechnology.htm>

OECD. (2017). *Consensus document on the biology of atlantic salmon (Salmo satar)*. <https://doi.org/https://doi.org/10.1787/9789264279728-en>

OECD. (2018). *Safety Assessment of Transgenic Organisms in the Environment, Volume 8 OECD Consensus Document of the Biology of Mosquito Aedes aegypti*. OECD. <https://doi.org/10.1787/9789264302235-en>

Oye, K. A., Esvelt, K., Appleton, E., Catteruccia, F., Church, G., Kuiken, T., Lightfoot, S. B.-Y., McNamara, J., Smidler, A., & Collins, J. P. (2014). Regulating gene drives. *Science*, 345(6197), 626–628. <https://doi.org/10.1126/science.1254287>

Rabitz, F. (2019). Gene drives and the international biodiversity regime. *Review of European, Comparative & International Environmental Law*, 28(3), 339–348. <https://doi.org/10.1111/reel.12289>

Redford, K. H., Brooks, T. M., Macfarlane, N. B. W., & Adams, J. S. (Eds.). (2019). *Genetic frontiers for conservation: an assessment of synthetic biology and biodiversity conservation: technical assessment*. IUCN, International Union for Conservation of Nature. <https://doi.org/10.2305/IUCN.CH.2019.05.en>

Reynolds, J. L. (2019). Governing New Biotechnologies for Biodiversity Conservation: Gene Drives, International Law, and Emerging Politics. *SSRN Electronic Journal*. <https://doi.org/10.2139/ssrn.3471735>

Rüdelsheim, P., & Smets, G. (2018). *Gene Drives: Experience with gene drive systems that may inform an environmental risk assessment.* <https://cogem.net/app/uploads/2019/07/CGM-2018-03-Report-Gene-Drives-met-kaft1.pdf>

Singh, J. A. (2019). Informed consent and community engagement in open field research: lessons for gene drive science. *BMC Medical Ethics*, 20(1), 54. <https://doi.org/10.1186/s12910-019-0389-3>

The Royal Society. (2018). *Gene drive research: why it matters.* <https://royalsociety.org/topics-policy/publications/2018/gene-drive-statement/>

Tourangeau, W. (2017). GMO doublespeak: An analysis of power and discourse in Canadian debates over agricultural biotechnology. *Canadian Food Studies / La Revue Canadienne Des Études Sur l'alimentation*, 4(1), 108. <https://doi.org/10.15353/cfs-rcea.v4i1.208>

van der Vlugt, C., van den Akker, H., Roesink, C., & Westra, J. (2018). *Risk assessment method for activities involving organisms with a gene drive under contained use.* <https://doi.org/10.21945/RIVM-2018-0090>

Webber, B. L., Raghu, S., & Edwards, O. R. (2015). Opinion: Is CRISPR-based gene drive a biocontrol silver bullet or global conservation threat? *Proceedings of the National Academy of Sciences*, 112(34), 10565–10567. <https://doi.org/10.1073/pnas.1514258112>

Westra, J., van der Vlugt, C., Roesink, C., Hogervorst, P., & Glandorf, D. (2016). *Gene Drives: Policy Report.* <https://www.rivm.nl/bibliotheek/rapporten/2016-0023.pdf>

Other relevant information

Böhne, A., Wilson, C. A., Postlethwait, J. H., & Salzburger, W. (2016). Variations on a theme: Genomics of sex determination in the cichlid fish *Astatotilapia burtoni*. *BMC Genomics*, 17(1), 883. <https://doi.org/10.1186/s12864-016-3178-0>

Eloranta, A. P., Johnsen, S. I., Power, M., Bærum, K. M., Sandlund, O. T., Finstad, A. G., Rognerud, S., & Museth, J. (2019). Introduced European smelt (*Osmerus eperlanus*) affects food web and fish community in a large Norwegian lake. *Biological Invasions*, 21(1), 85–98. <https://doi.org/10.1007/s10530-018-1806-0>

Gemmell, N. J., Todd, E. V., Goikoetxea, A., Ortega-Recalde, O., & Hore, T. A. (2019). *Natural sex change in fish* (pp. 71–117). <https://doi.org/10.1016/bs.ctdb.2018.12.014>

Ghalambor, C. K., Hoke, K. L., Ruell, E. W., Fischer, E. K., Reznick, D. N., & Hughes, K. A. (2015). Non-adaptive plasticity potentiates rapid adaptive evolution of gene expression in nature. *Nature*, 525(7569), 372–375. <https://doi.org/10.1038/nature15256>

Glover, K. A., Solberg, M. F., McGinnity, P., Hindar, K., Verspoor, E., Coulson, M. W., Hansen, M. M., Araki, H., Skaala, Ø., & Svåsand, T. (2017). Half a century of genetic interaction between farmed and wild Atlantic salmon: Status of knowledge and unanswered questions. *Fish and Fisheries*, 18(5), 890–927. <https://doi.org/10.1111/faf.12214>

Hill, J. E., Tuckett, Q. M., Hardin, S., Lawson, L. L., Lawson, K. M., Ritch, J. L., & Partridge, L. (2017). Risk Screen of Freshwater Tropical Ornamental Fishes for the Conterminous United States. *Transactions of the American Fisheries Society*, 146(5), 927–938. <https://doi.org/10.1080/00028487.2017.1312523>

Hu, L., Huang, M., Tang, M., Yu, J., & Zheng, B. (2015). Wolbachia spread dynamics in stochastic environments. *Theoretical Population Biology*, 106, 32–44. <https://doi.org/10.1016/j.tpb.2015.09.003>

Jensen, A., Karlsson, S., Fiske, P., Hansen, L., Hindar, K., & Østborg, G. (2013). Escaped farmed Atlantic salmon grow, migrate and disperse throughout the Arctic Ocean like wild salmon. *Aquaculture Environment Interactions*, 3(3), 223–229. <https://doi.org/10.3354/aei00064>

Jiggins, F. M. (2017). The spread of Wolbachia through mosquito populations. *PLOS Biology*, 15(6), e2002780. <https://doi.org/10.1371/journal.pbio.2002780>

Lam, M. E. (2016). The Ethics and Sustainability of Capture Fisheries and Aquaculture. *Journal of Agricultural and Environmental Ethics*, 29(1), 35–65. <https://doi.org/10.1007/s10806-015-9587-2>

Oomen, R. A., & Hutchings, J. A. (2015). Genetic variability in reaction norms in fishes. *Environmental Reviews*, 23(3), 353–366. <https://doi.org/10.1139/er-2014-0077>

Todd, E. V., Liu, H., Muncaster, S., & Gemmell, N. J. (2016). Bending Genders: The Biology of Natural Sex Change in Fish. *Sexual Development*, 10(5–6), 223–241. <https://doi.org/10.1159/000449297>

Wellband, K. W., & Heath, D. D. (2017). Plasticity in gene transcription explains the differential performance of two invasive fish species. *Evolutionary Applications*, 10(6), 563–576. <https://doi.org/10.1111/eva.12463>
